



***Proteomics
Standards Research
Group (sPRG)***

sPRG: a Tale of Two Studies

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sPRG working group members

Toni Koller (Acting Chair)	Columbia University
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Christopher Colangelo	Primary Ion
David Hawke	UT MD Anderson Cancer Center
Alexander R. Ivanov	Northeastern University
Gordana Ivosev	Sciex
Paul Rudnick	Spectragen Informatics
Brian C. Searle	Proteome Software / U. of Washington
Scott A. Shaffer	U. of Massachusetts Medical School

sPRG working group goals

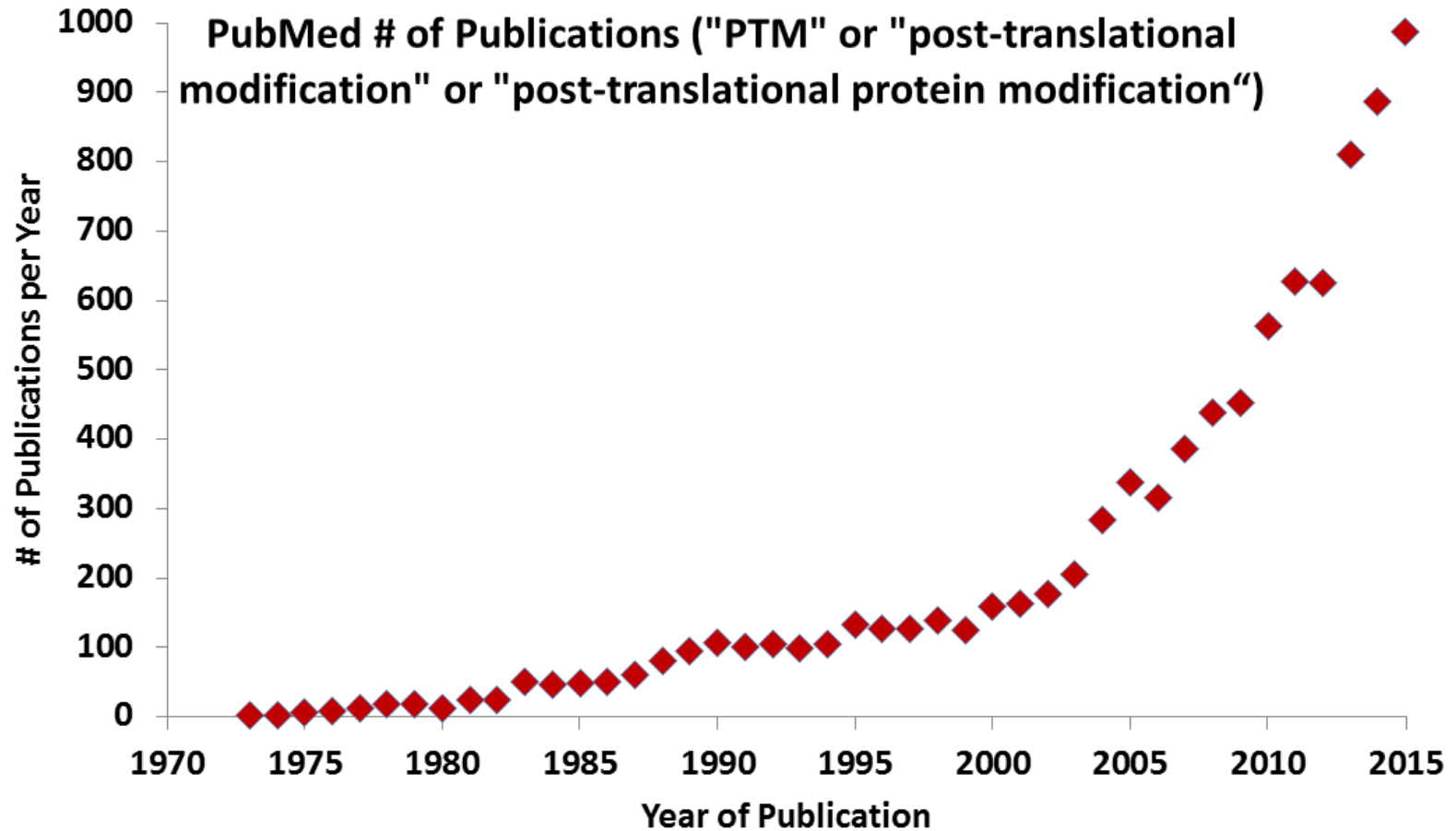
- Revise interpretations of previous studies
- Prepare manuscripts
- Make ABRF standards available to the community



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Revisiting the sPRG 2012 PTM study

PTMs continue to be a growing interest to proteomics



PRG 2003

- 2 digested proteins
- 2 synthetic phosphopeptides

Results:

- 54 labs returned data sets
- 5 identified 1 phosphopeptide
- 5 identified the other
- 3 identified both
- **Massive over reporting**

sPRG 2007

- Mixture of 7 phosphorylated proteins

Results:

- 44 labs returned data sets
- 50 “known” sites of phosphorylation
- 27 sites identified by multiple labs
- 8 “bonus” sites identified by multiple labs
- Only 5 sites identified by $\geq 50\%$ of labs
- **Over reporting? Interpretation hampered by unknowns**

sPRG 2010

- 6 digested proteins
- 23 synthetic phosphopeptides

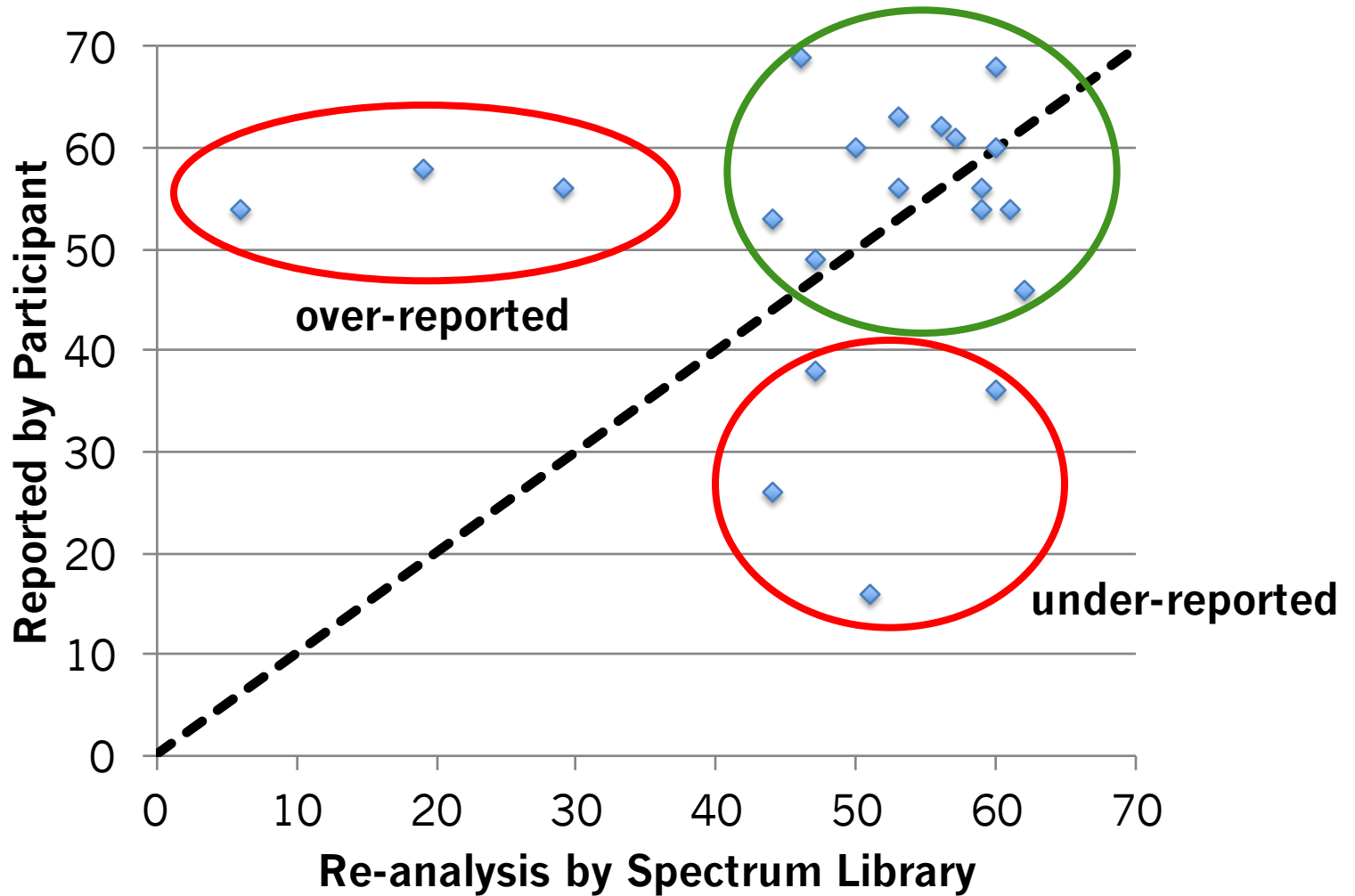
Results:

- 43 labs returned data sets
- 23 sites identified by multiple labs
- 16 sites identified by $\geq 50\%$ of labs
- Multiply phosphorylated peptides still a challenge

sPRG 2012

- 6 digested proteins
- 45 synthetic phosphopeptides
 - (including positional isomers)
- 41 synthetic modified peptides
 - sulfated tyrosine
 - nitrosylated tyrosine
 - acetylated lysine
 - mono- di- and tri-methylated arginine/lysine
 - sym/asymmetric di-methylated arginine
- 30 data sets returned

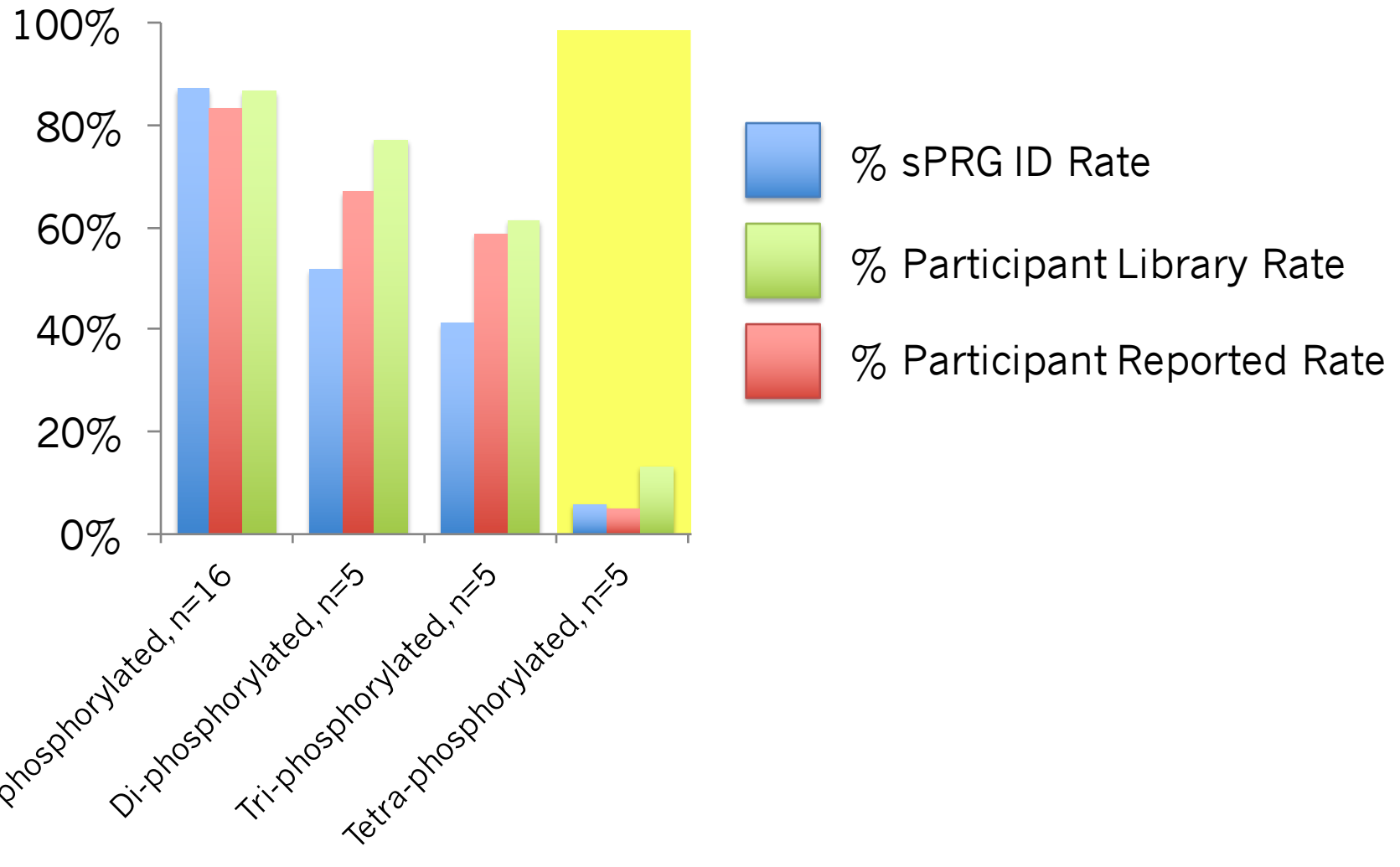
Over-reporting has been curbed somewhat



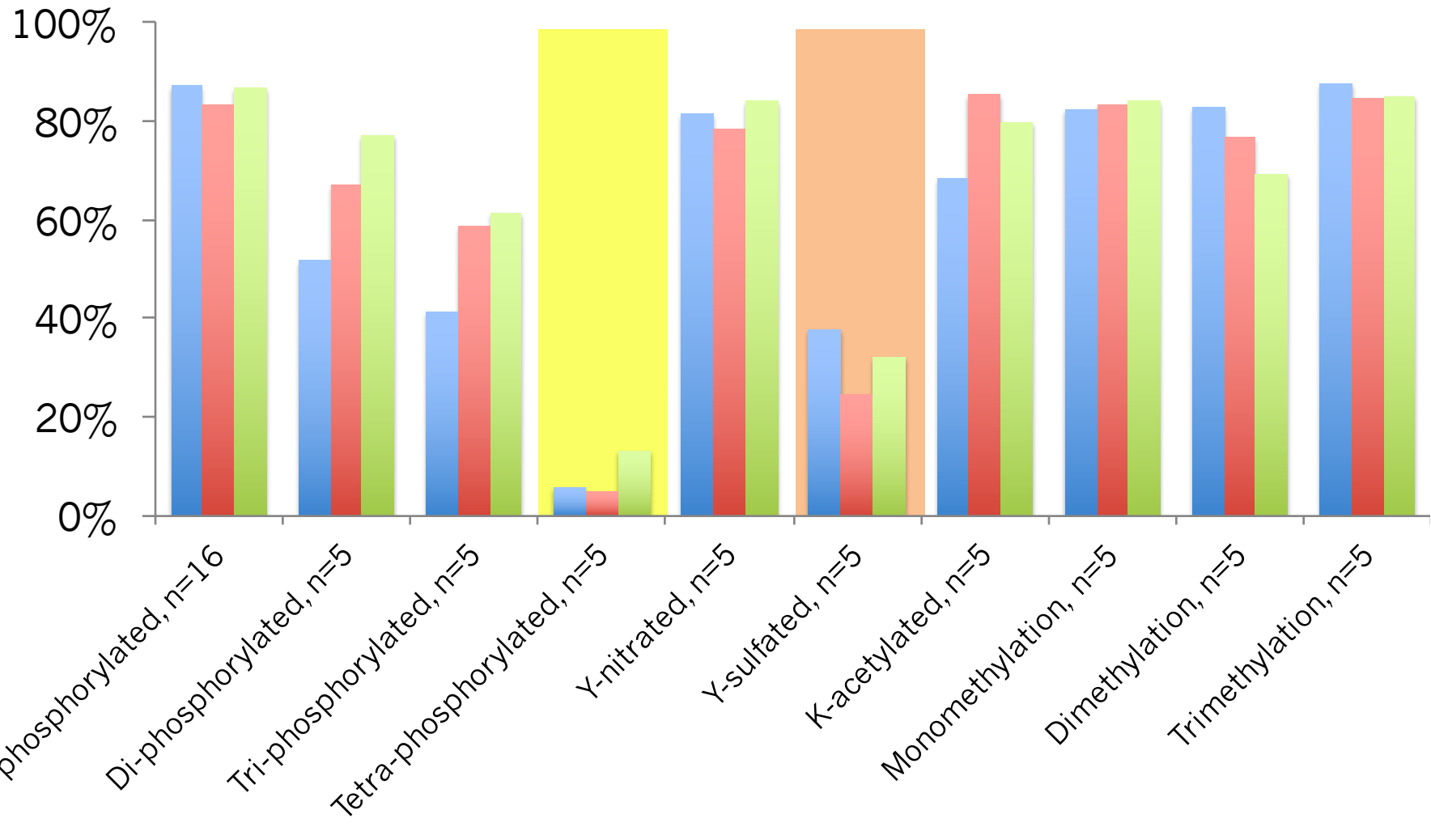
Cross study comparison shows general improvement

	PRG 2003	sPRG 2010	sPRG 2012
SVSpDYEGK	15%	40%	80%
THILLFLPKSpVSDYEGK	15%	62%	80%

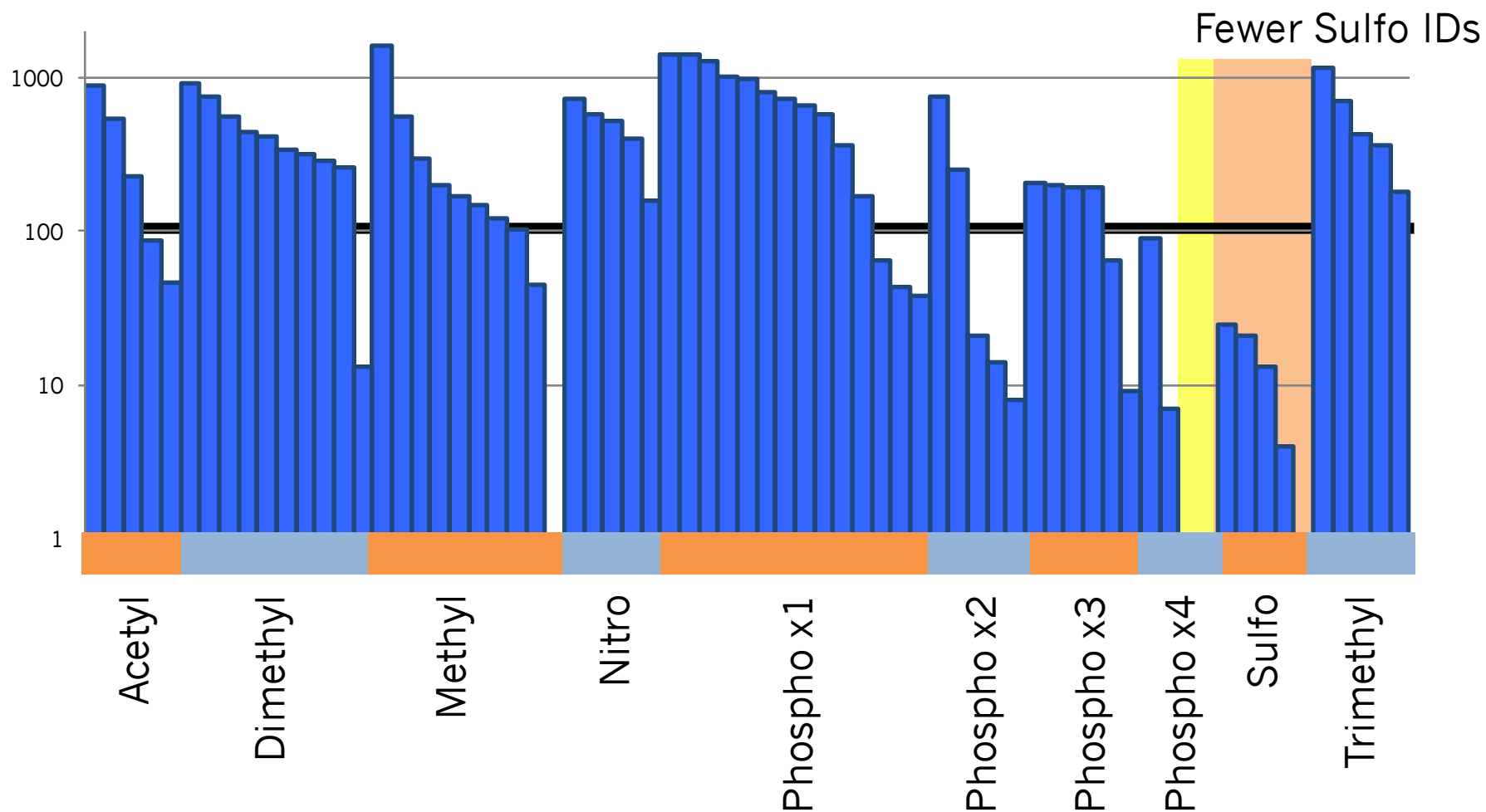
Still difficult to identify multiply phosphorylated peptides



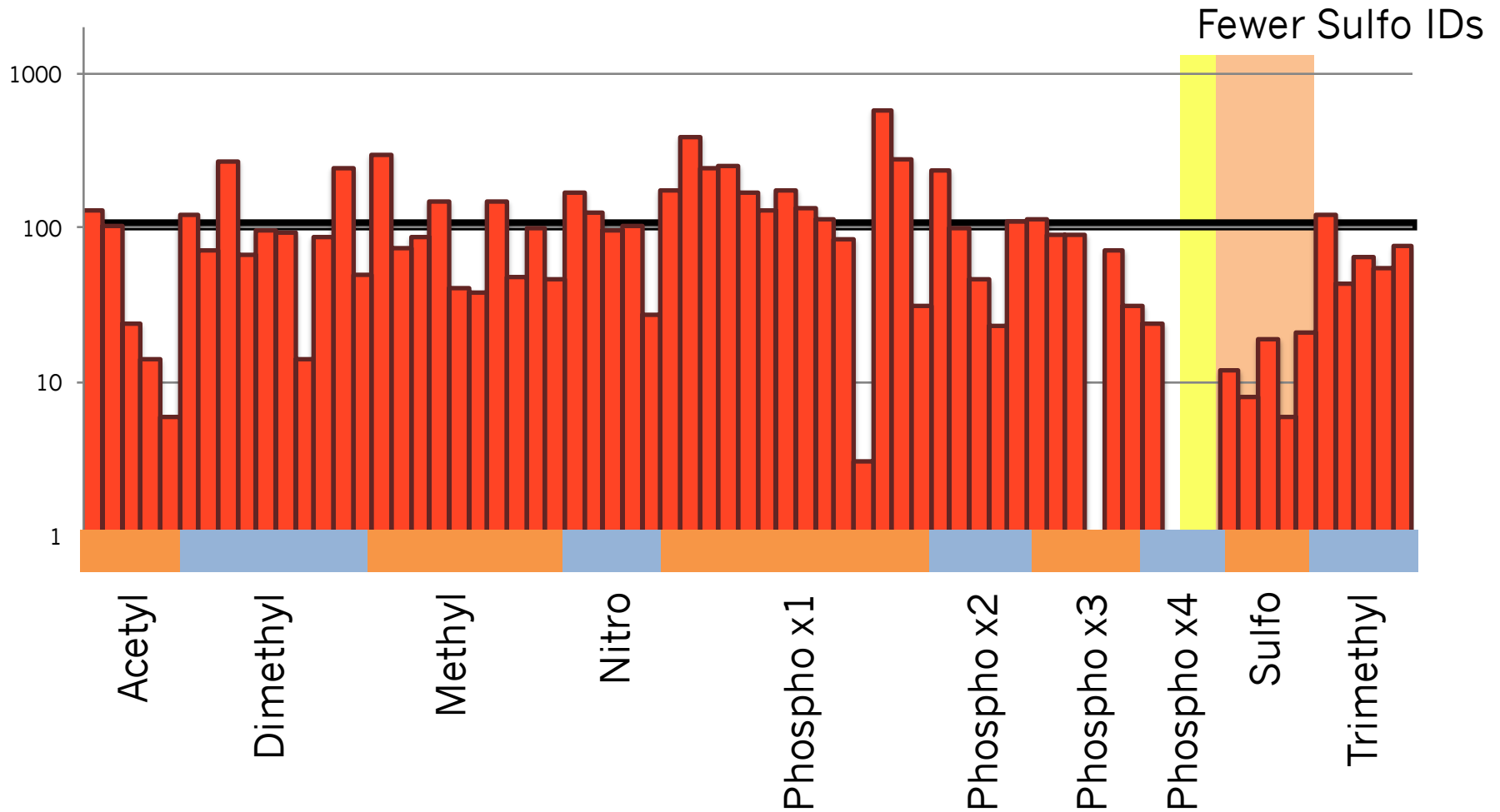
Analyses of other modifications are more successful



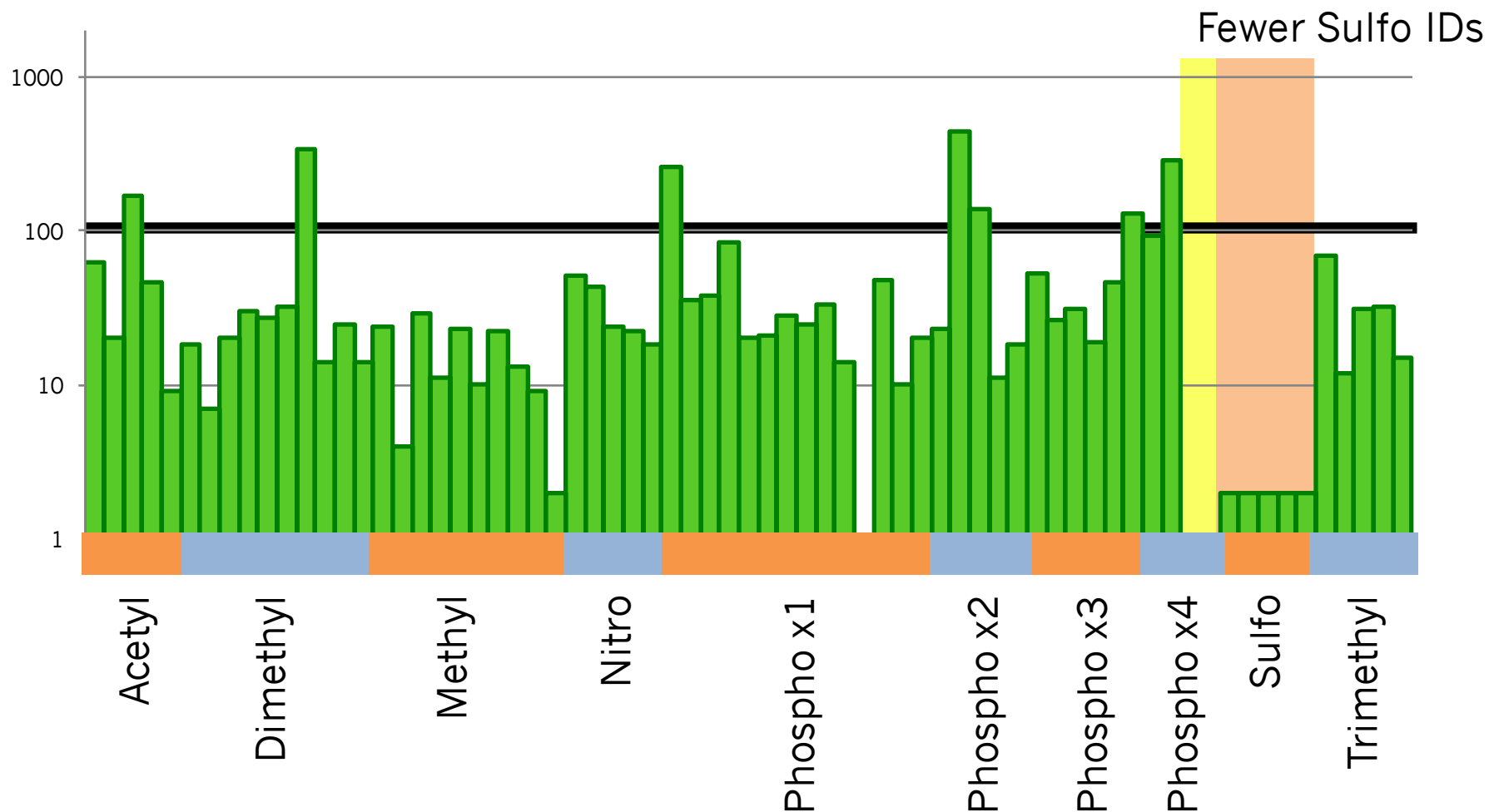
Confident CID identifications



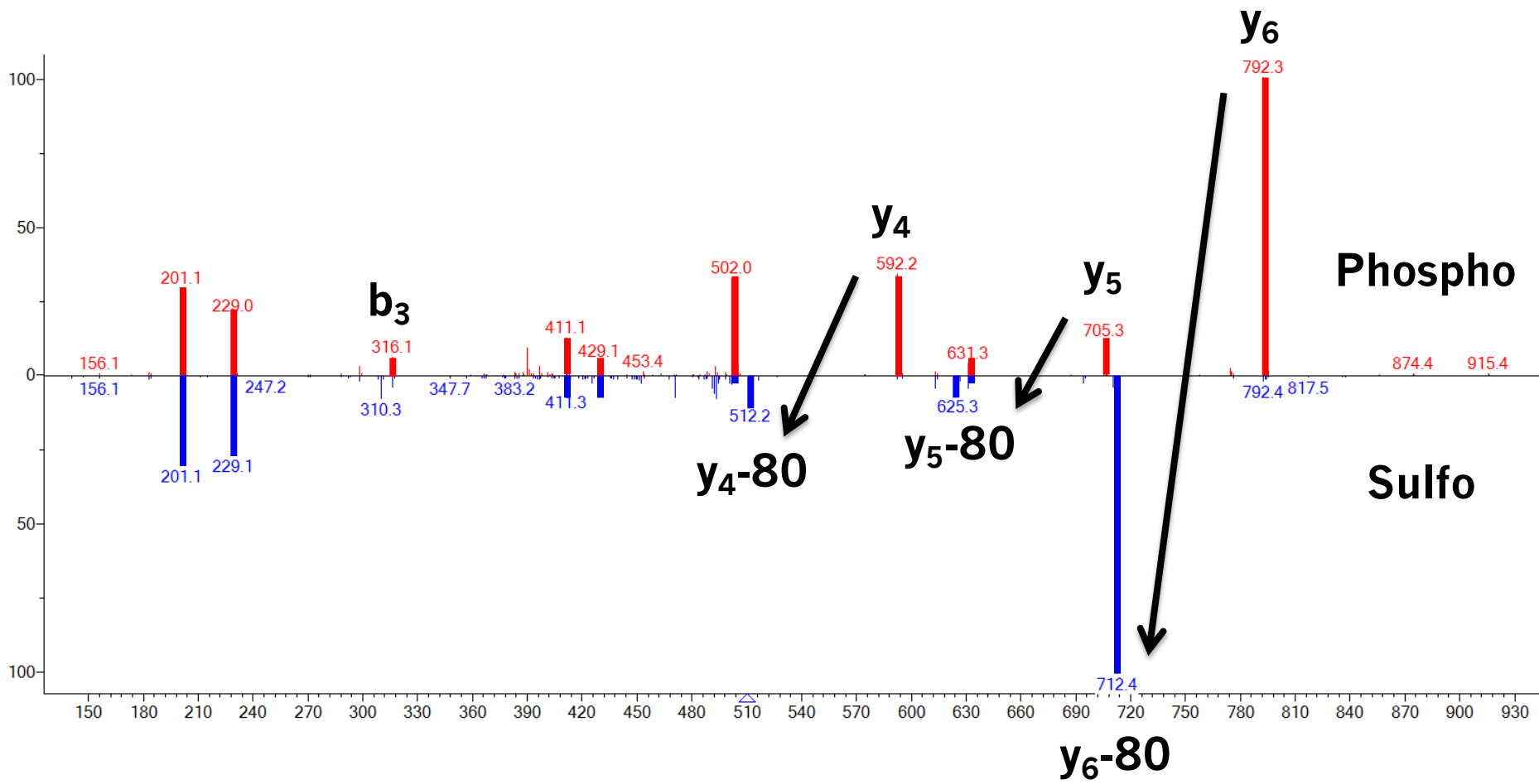
Confident HCD identifications



Confident Q-ToF identifications



DISL**S***DYK (Phospho/Sulfo)



Standard Availability

- Working with Thermo Fisher and Spectragen Informatics to distribute the sample with a new mass spectral library
- Revalidated the sample to confirm the make up
- Commercially available in limited quantities soon
- Sign up to be notified of its availability at <http://spectragen-informatics.com/sprg>

sPRG members involved in this study

Alexander R. Ivanov (Chair)	Northeastern University
Christopher Colangelo	Primary Ion
Craig Dufresne	Thermo Fisher Scientific
David Friedman	Vanderbilt University
Kathryn S. Lilley	University of Cambridge
Karl Mechtler	IMP Research Inst. of Molecular Pathology
Brett Phinney	University of California, Davis
Kristie Rose	Vanderbilt University
Paul Rudnick	Spectragen Informatics
Brian C. Searle	Proteome Software / U. of Washington
Scott A. Shaffer	U. of Massachusetts Medical School
Susan T. Weintraub	University of Texas HSC

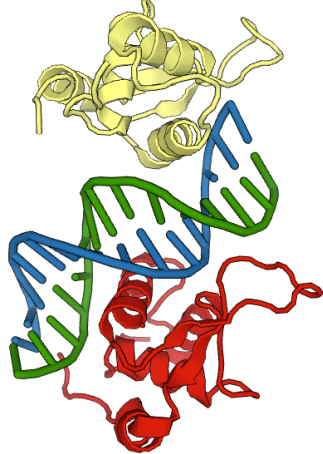


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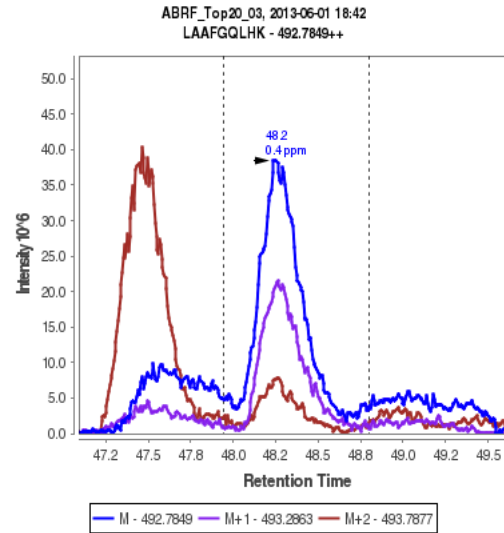
Revisiting the sPRG 2014 “1000 Peptide” quantification study

Relative Quantification with Stable Isotope Labeled Peptides

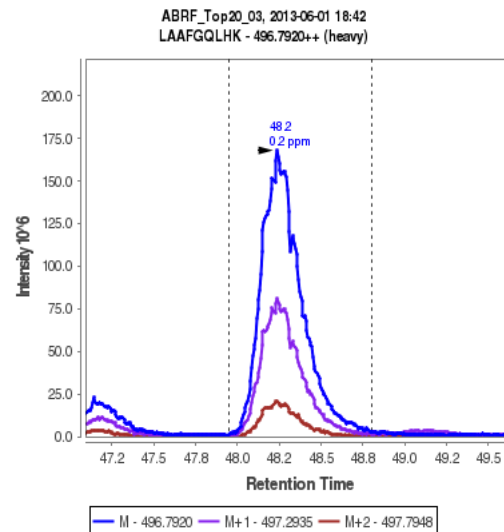
ILF3 (interleukin enhancer binding factor 3)



LAAF¹³FGQLHK



LAAF¹³FGQLHK*



light

heavy



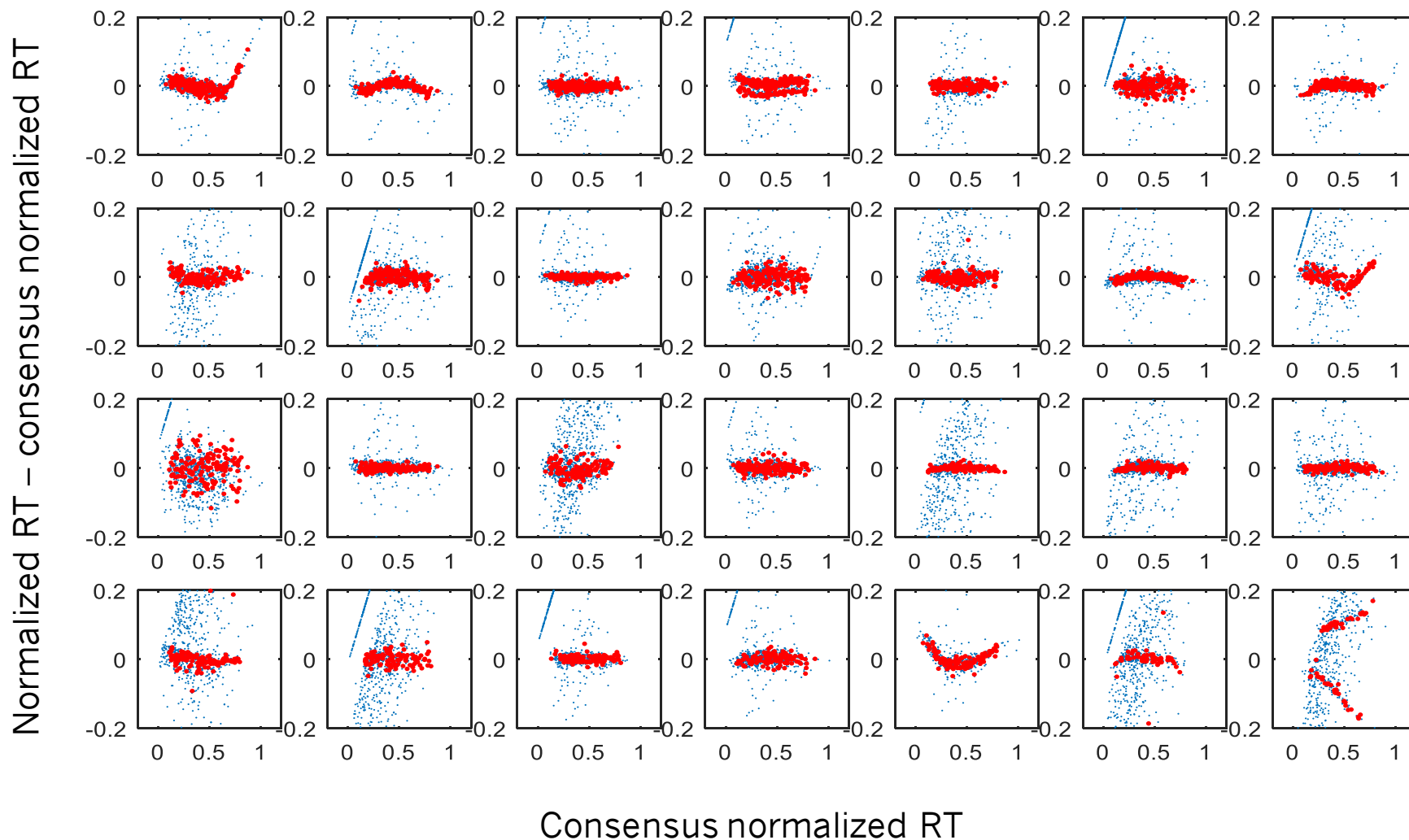
sPRG 2014

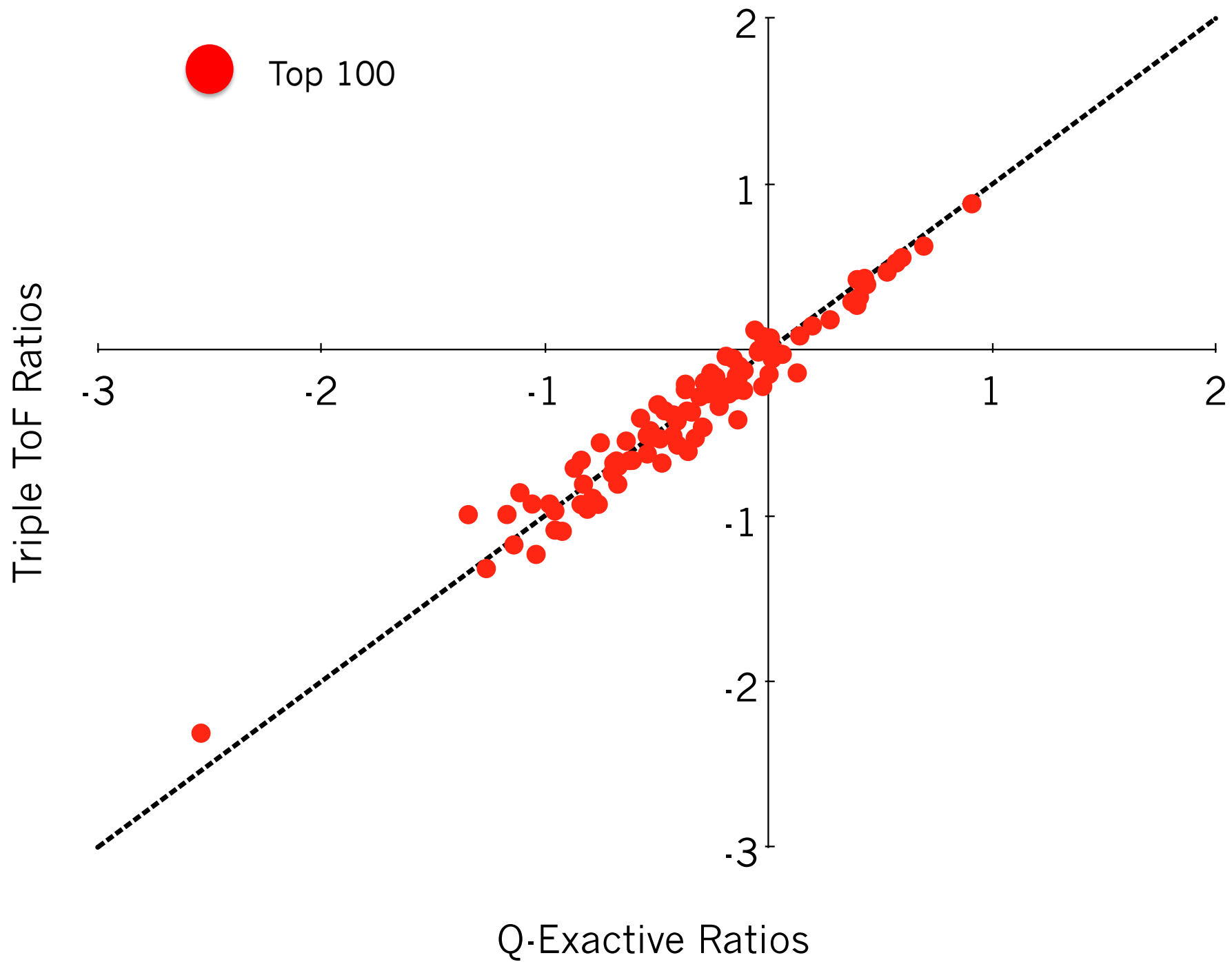
- 1000 tryptic peptides from 552 proteins synthesized by JPT
- Conserved across *Homo sapiens*, *Mus musculus* and *Rattus norvegicus*
- Chosen because of consistency of observation across three different labs
- stable isotope labeled at R and K

sPRG 2014

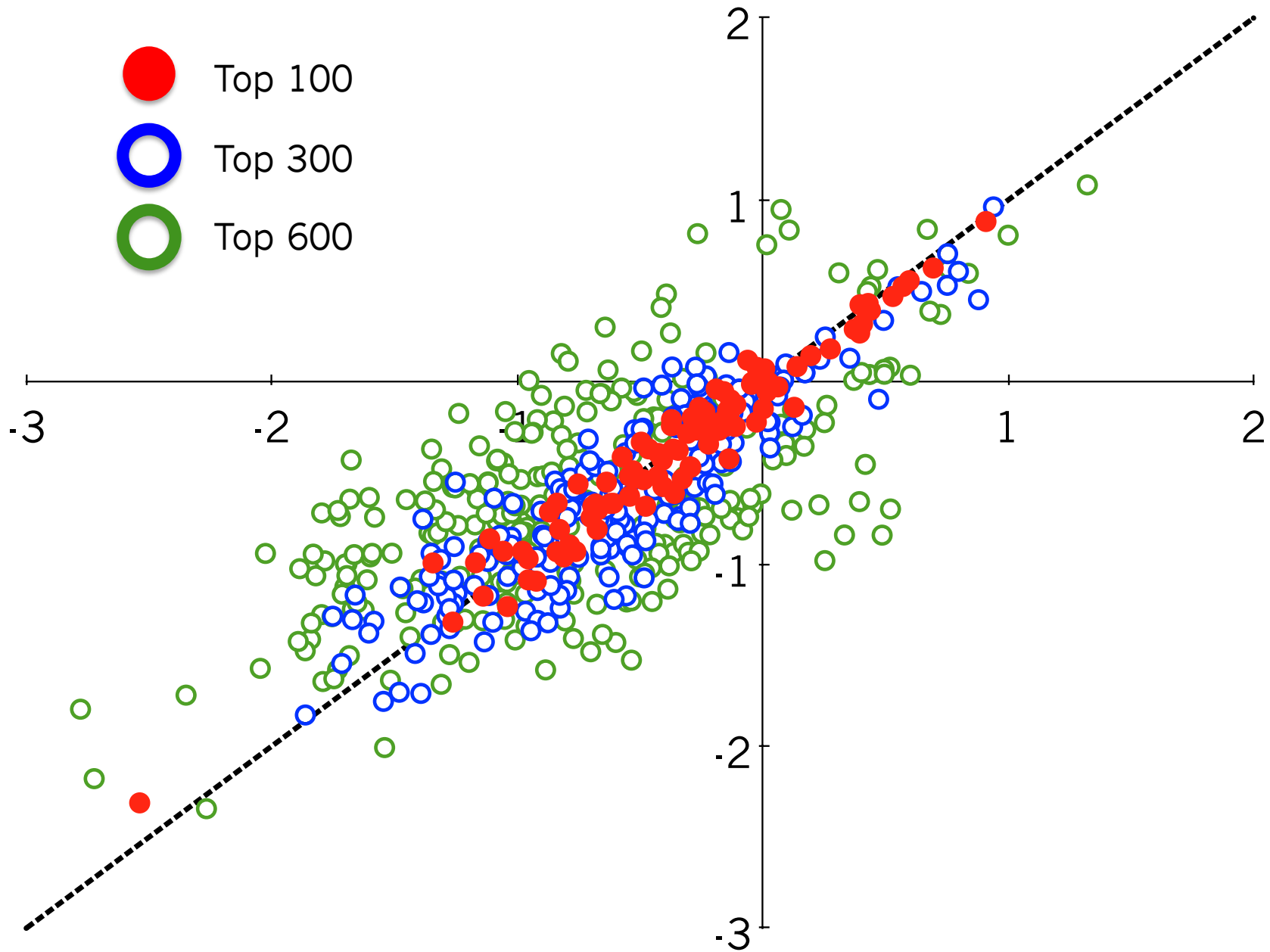
- Only light cleanup: we don't know the true abundance of the peptides
- When mixed with other samples: provides a relative standard to compare across platforms
- Initial study performed with HEK 293 matrix
- 49 labs returned data sets

Retention times are extremely consistent across labs and platforms





Triple ToF Ratios

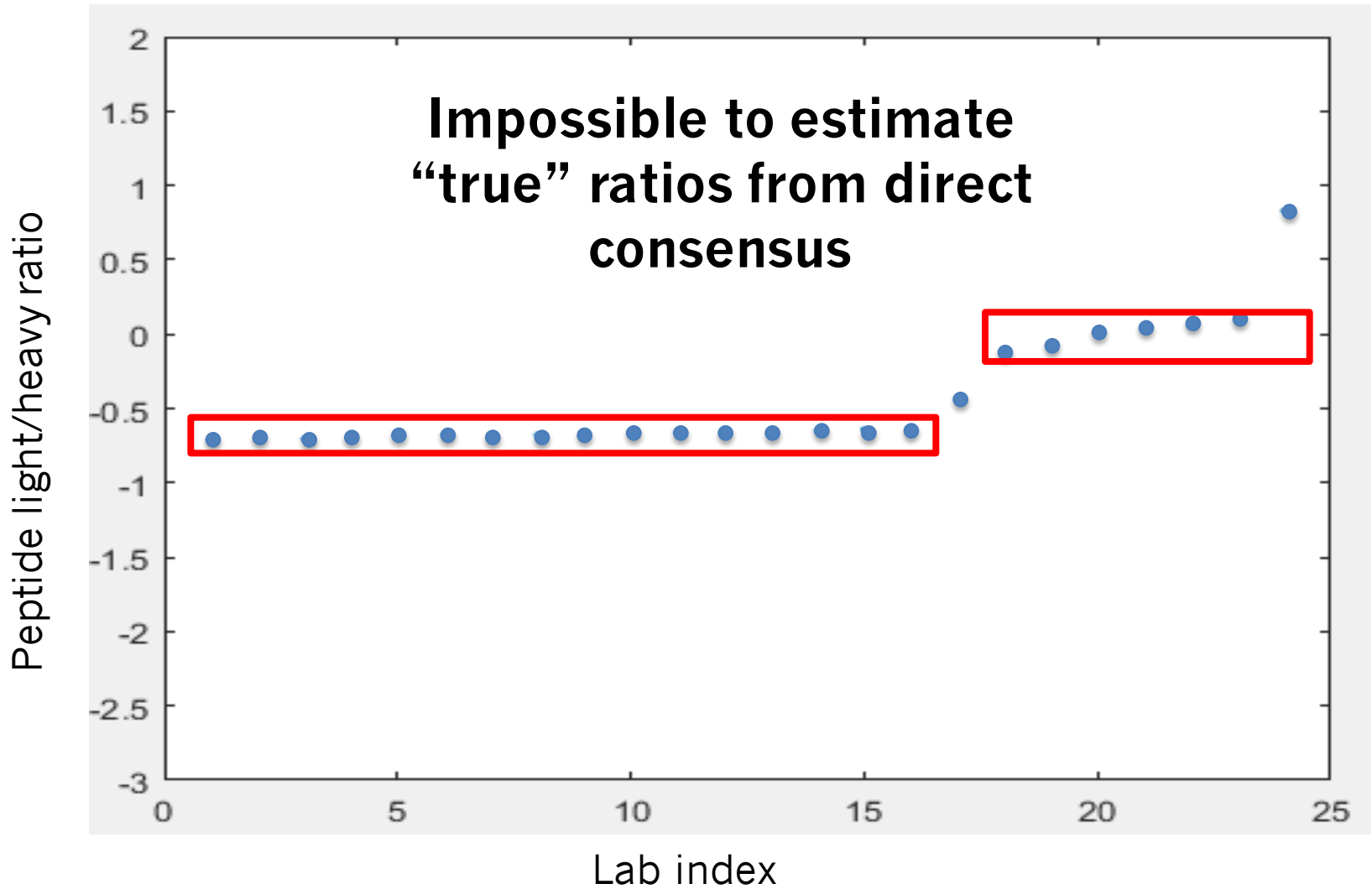


Q-Exactive Ratios

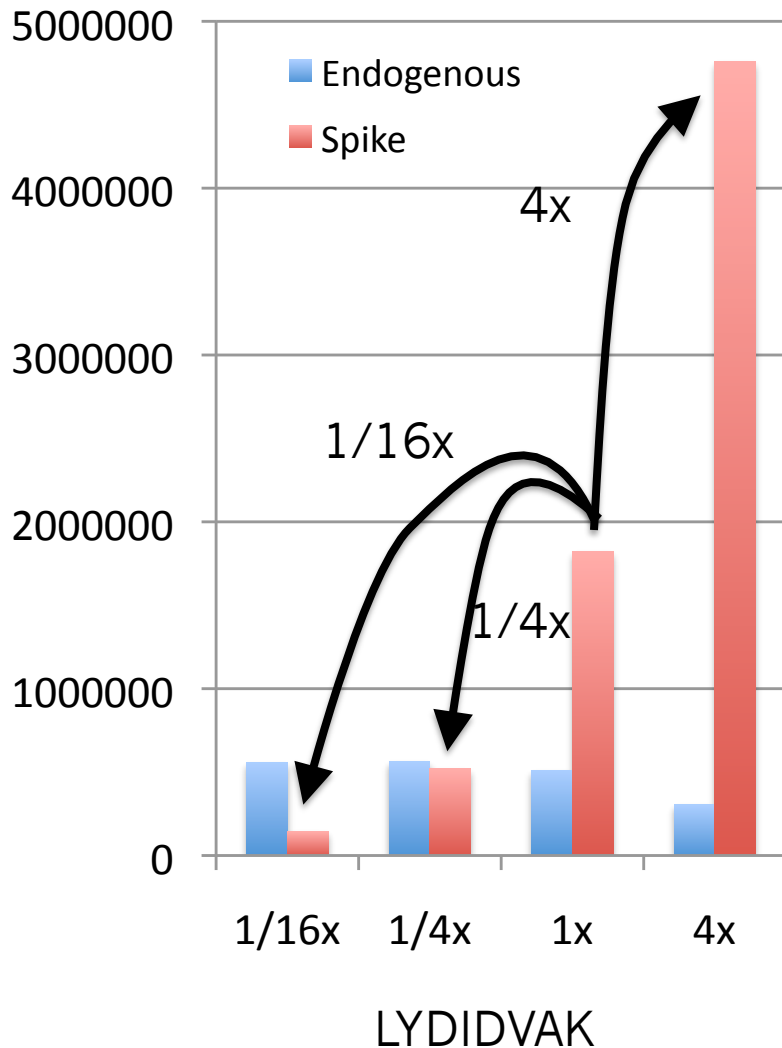
Much more quantitative variability than we expected!

- Worked to assign a better “true” ratio to improve alignment
- Worked to understand where the variability was coming from

What is the “true” peptide ratio?

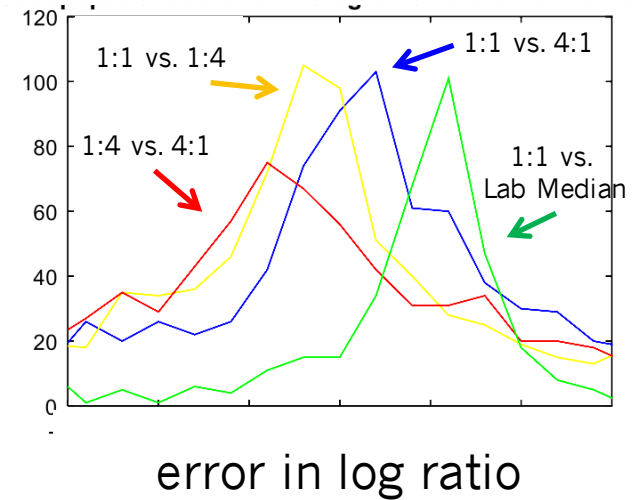
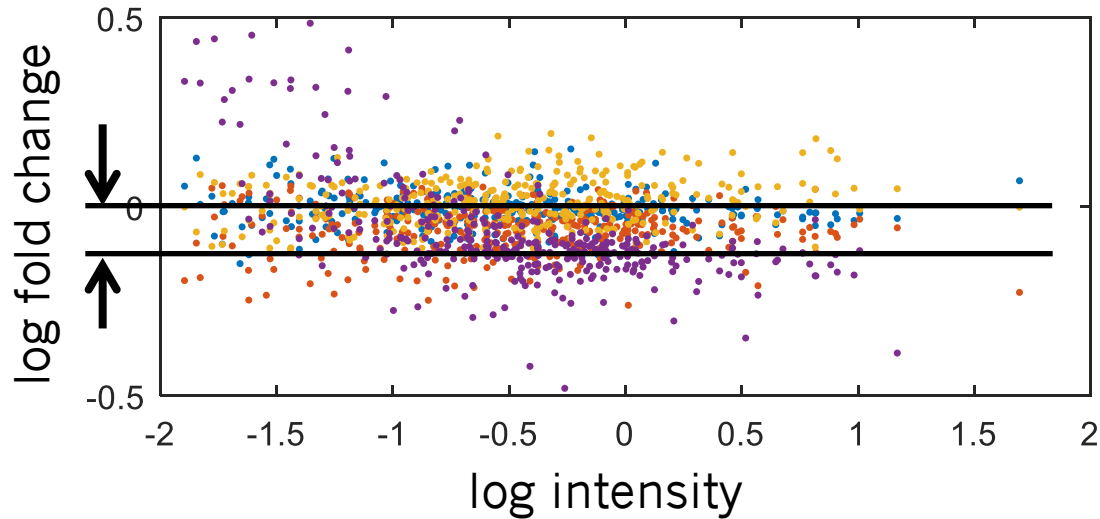


Constructing an accurate “true” ratio

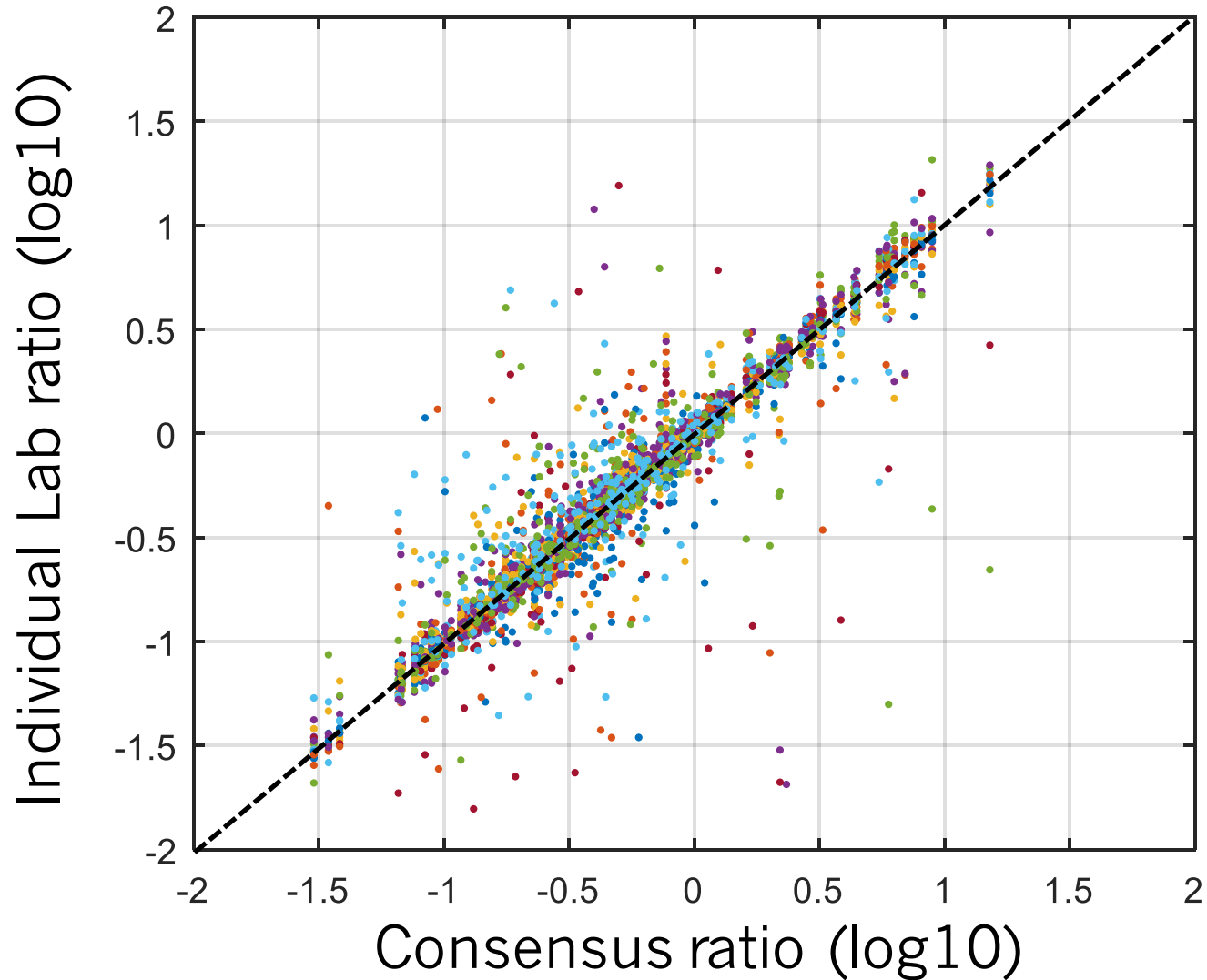


- Only 1x was given to participants
- Triplicate analysis of all mixtures
- Two very different instruments / configurations

Assigning a “true” ratio from all the dilution mixtures



357 peptides in good agreement across most labs



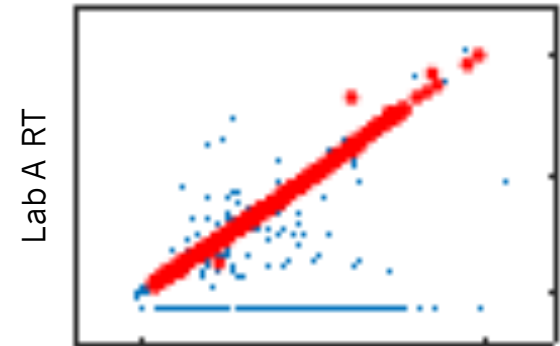
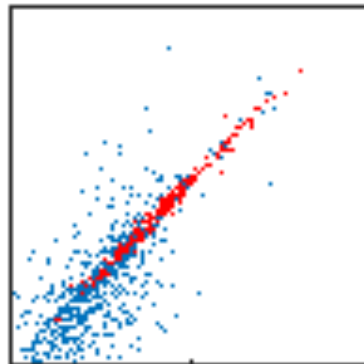
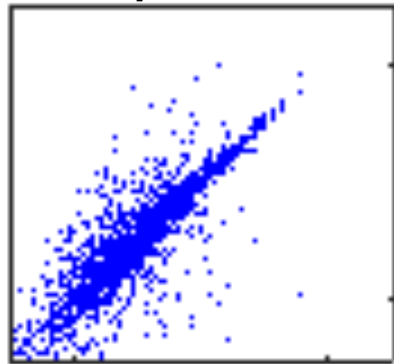
Replicate accuracy doesn't necessarily imply "true" accuracy

Ratio agreement within technical replicates

Ratio agreement with consensus

RT agreement with consensus

Lab A
Technical
Replicate Ratio 1

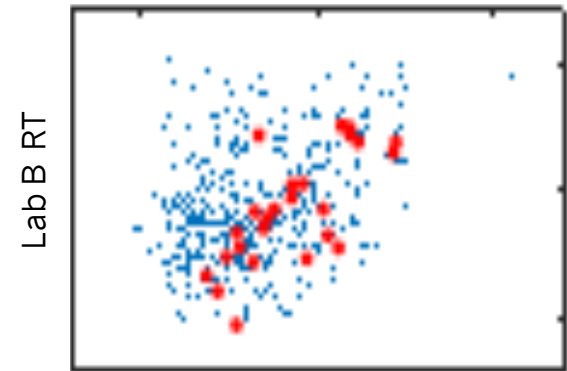
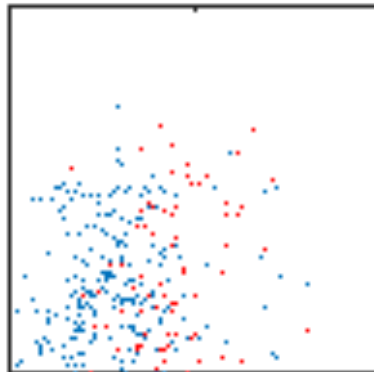
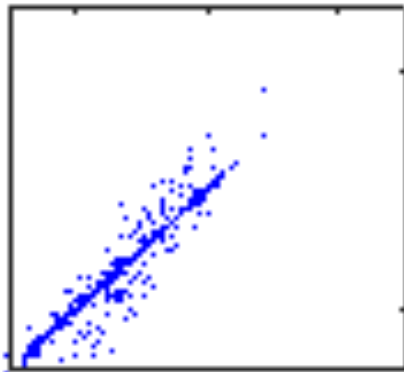


Technical
Replicate Ratio 2

Consensus Ratio

Consensus RT

Lab B
Technical
Replicate Ratio 1

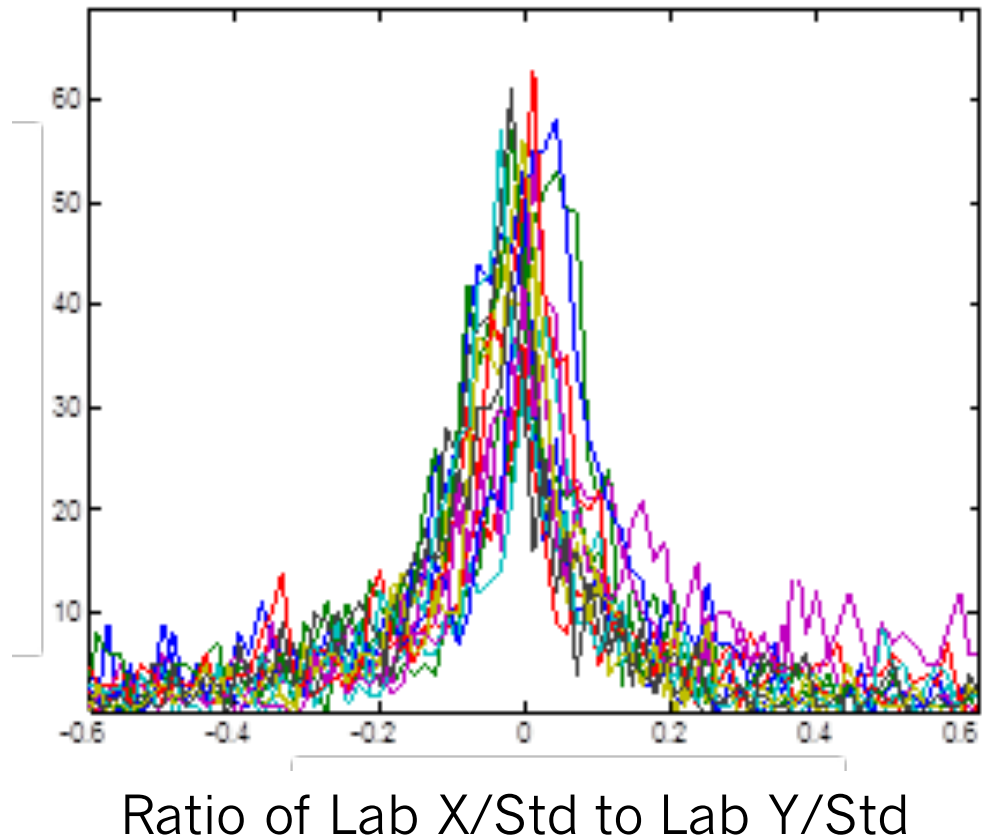
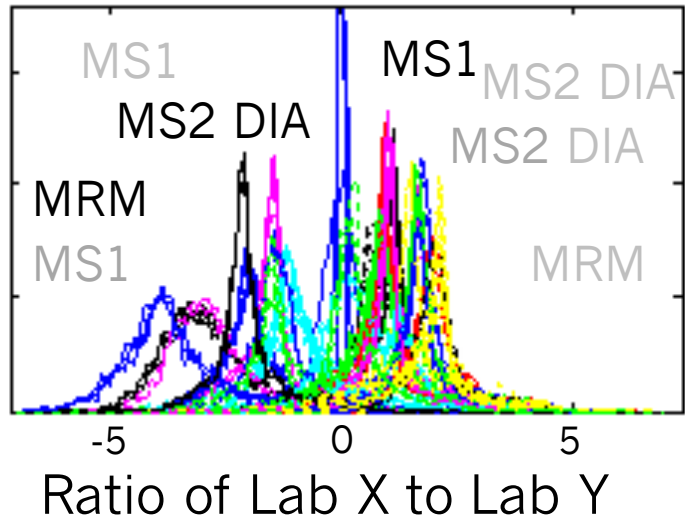


Technical
Replicate Ratio 2

Consensus Ratio

Consensus RT

Adding standard to sample allows comparison to other labs/platforms



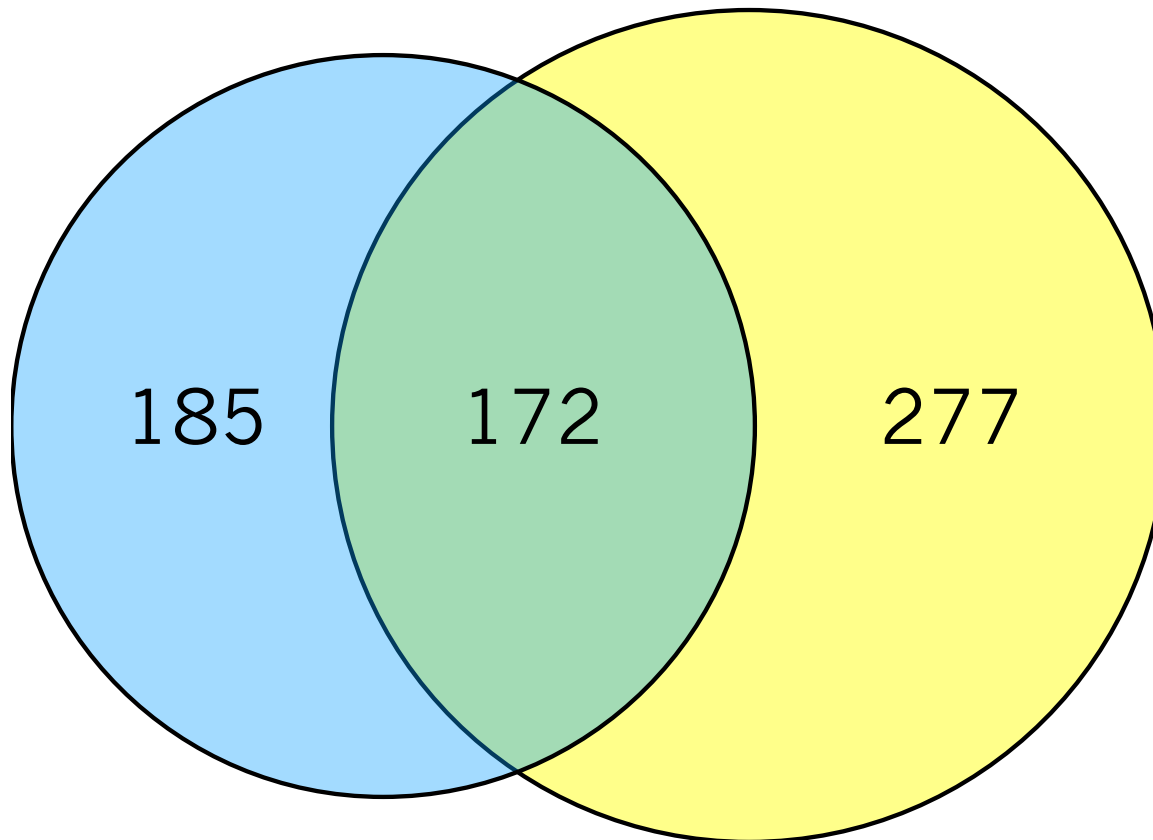
Re-characterized standard mixed with in HeLa

- Logically, if you can compare very disparate platforms, you should be able to compare cell lines
- We ran acquisitions on 3x different instrument platforms

Different 300 peptides in HeLa?!

357 consensus
HEK peptide ratios

449 confidently IDed
HeLa peptide ratios



What does that mean? Take homes

- Matrix complexity has a huge effect on which peptides are visible
- “1000 peptide” standard doesn’t mean 1000 peptides are quantitative in your sample
- 1000 peptides sounds like overkill but it guarantees some peptides are quantitative
- Of the 1000 peptides, we believe approximately $1/3^{\text{rd}}$ are quantitative in any given cell line

What can you use this sample for if you don't have multiple platforms?

- Costs ~ 50¢ per sample (50 fmol)
- Cheap quantitative standard (if it overlaps with your peptides of interest)
- Loading standard
- iRT alignment standard for improving identification rates



Innovative Peptide Solutions

Proteomics & Enzyme Profiling

The advertisement graphic for SpikeMix™ features a background of laboratory petri dishes. On the left, there is a 3D ribbon diagram of a protein structure in shades of green and blue, with a red chromatogram line overlaid. The text 'SpikeMix™' is prominently displayed in a red, serif font, preceded by a decorative cluster of red dots. Below this, the text 'Ready-to-use proteotypic heavy-labeled peptide pools for mass spectrometry' is written in a smaller red font. A red starburst badge with the word 'NEW' in white is positioned to the right of the product name. The background also shows faint labels like 'DR474808', '809', '810', and 'DR474811'.

SpikeMix™ ABRF (cross-species standard)

100pmol - \$ 1049.00

10pmol - \$ 545.00

1pmol - \$ 164.00

sPRG members involved in this study

Christopher Colangelo (Chair) Primary Ion

Craig Dufresne

Thermo Fisher Scientific

David Hawke

UT MD Anderson Cancer Center

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and:

Brendan MacLean

U. of Washington

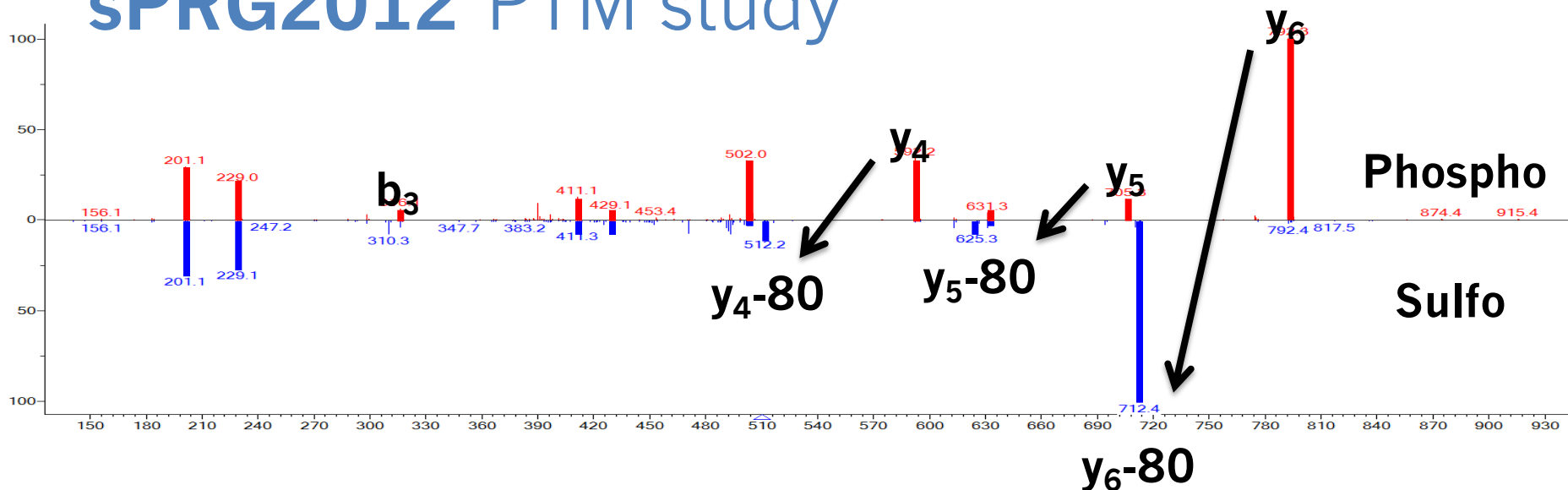
Vagisha Sharma

U. of Washington

sPRG 2017?

We have several new study ideas,
but need new members!

sPRG2012 PTM study



sPRG2014 1000 peptide study

