

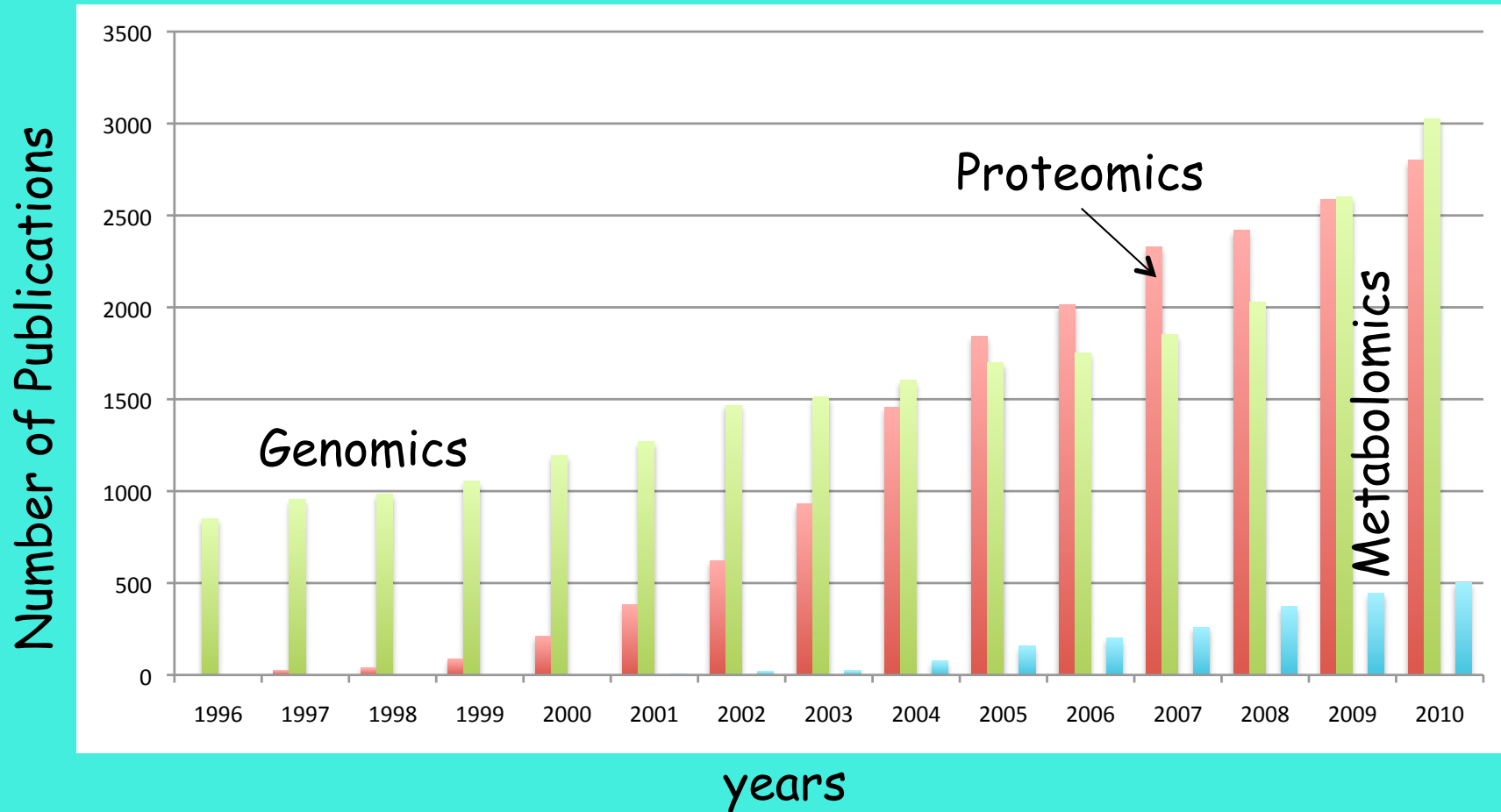
Pathway Analysis In Expression Proteomics

Roman Zubarev

Roman.Zubarev@ki.se

*Molecular Biometry,
Department for Medical Biochemistry & Biophysics,
Karolinska Institutet, Stockholm*

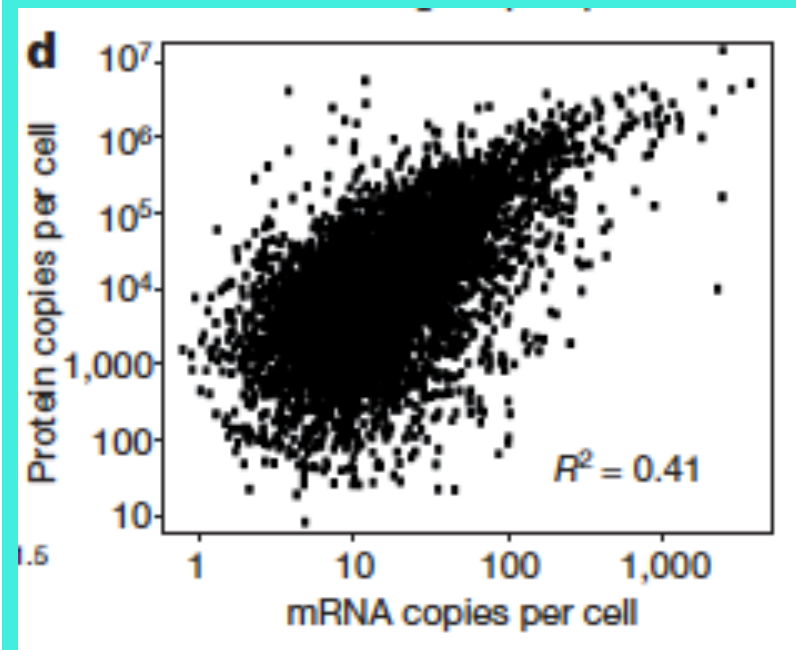
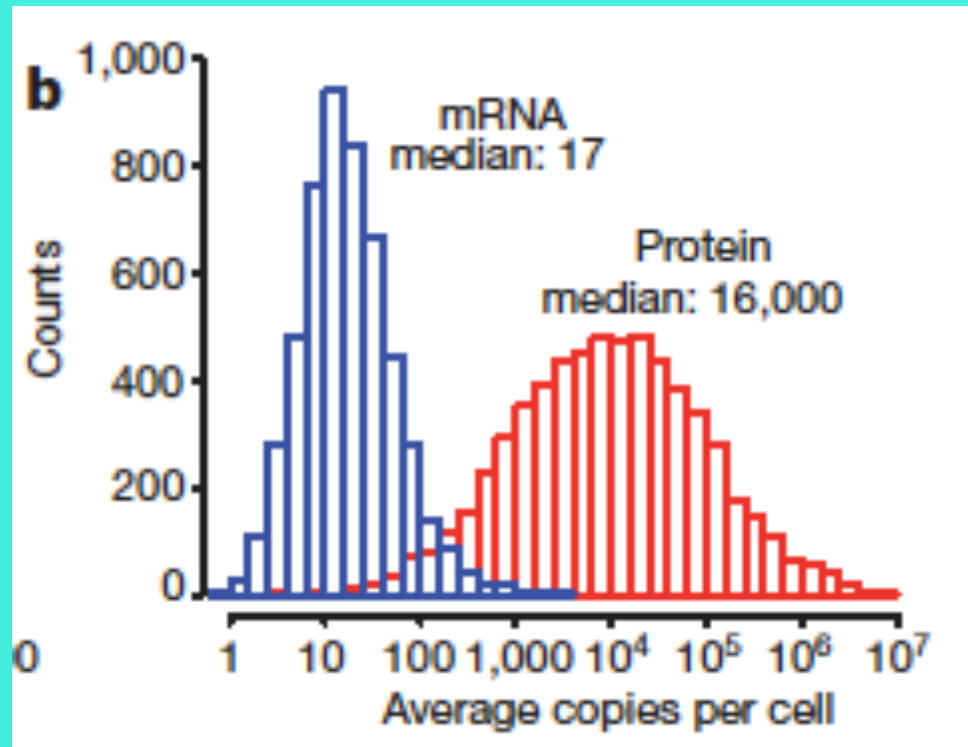
Proteomics vs Transcriptomics and Metabolomics



Genomics - what the cell *may* do
Transcriptomics - *wants* to do
Proteomics - *does*
Metabolomics - *has done*

Differences between transcriptomics and proteomics

- The dynamic range - 10^3 - 10^4 vs 10^7 .

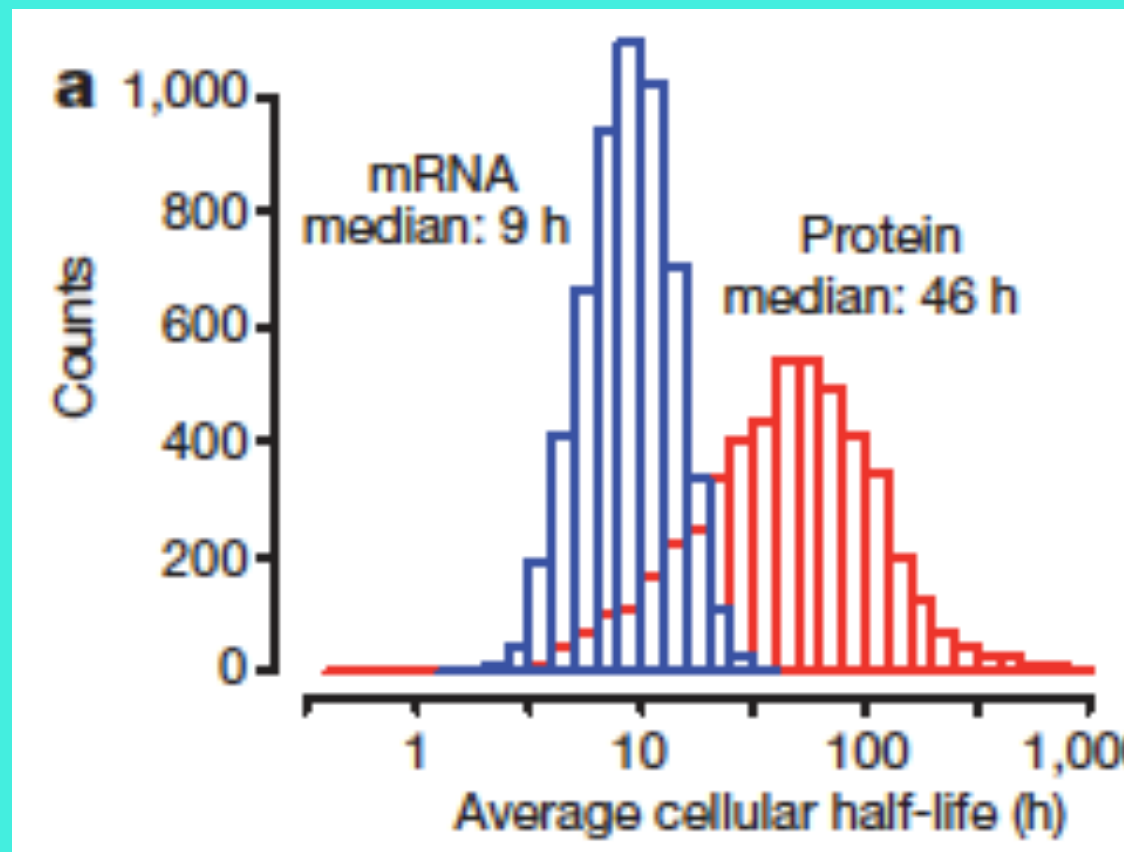


19 MAY 2011 | VOL 473 | NATURE | 337

Since the dynamic range of instrumentation is - 10^3 - 10^4 , transcriptomics easily covers all 10,000 expressed genes, while proteomics - ca. 5,000 proteins. But false discovery rate for mRNA 5%, for proteins - 1%

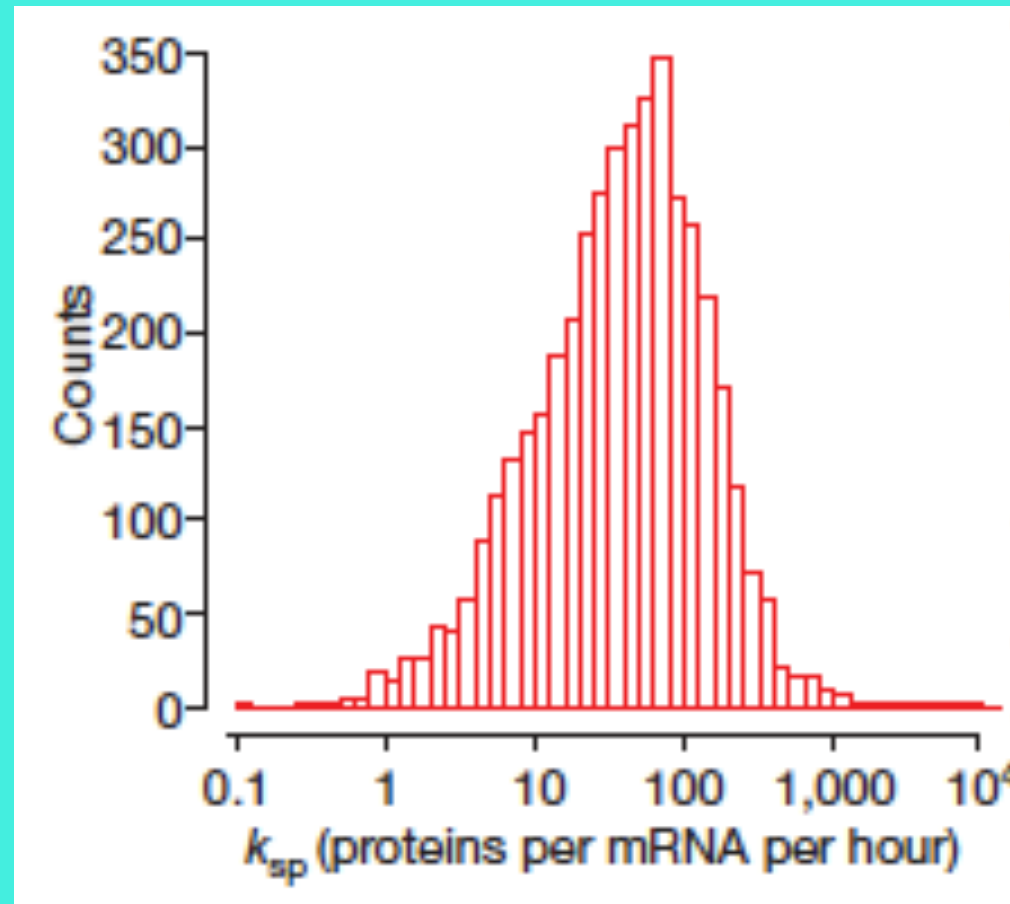
Differences between transcriptomics and proteomics

- The cellular half-life:
 - mRNA - 9h
 - proteins - 46 h

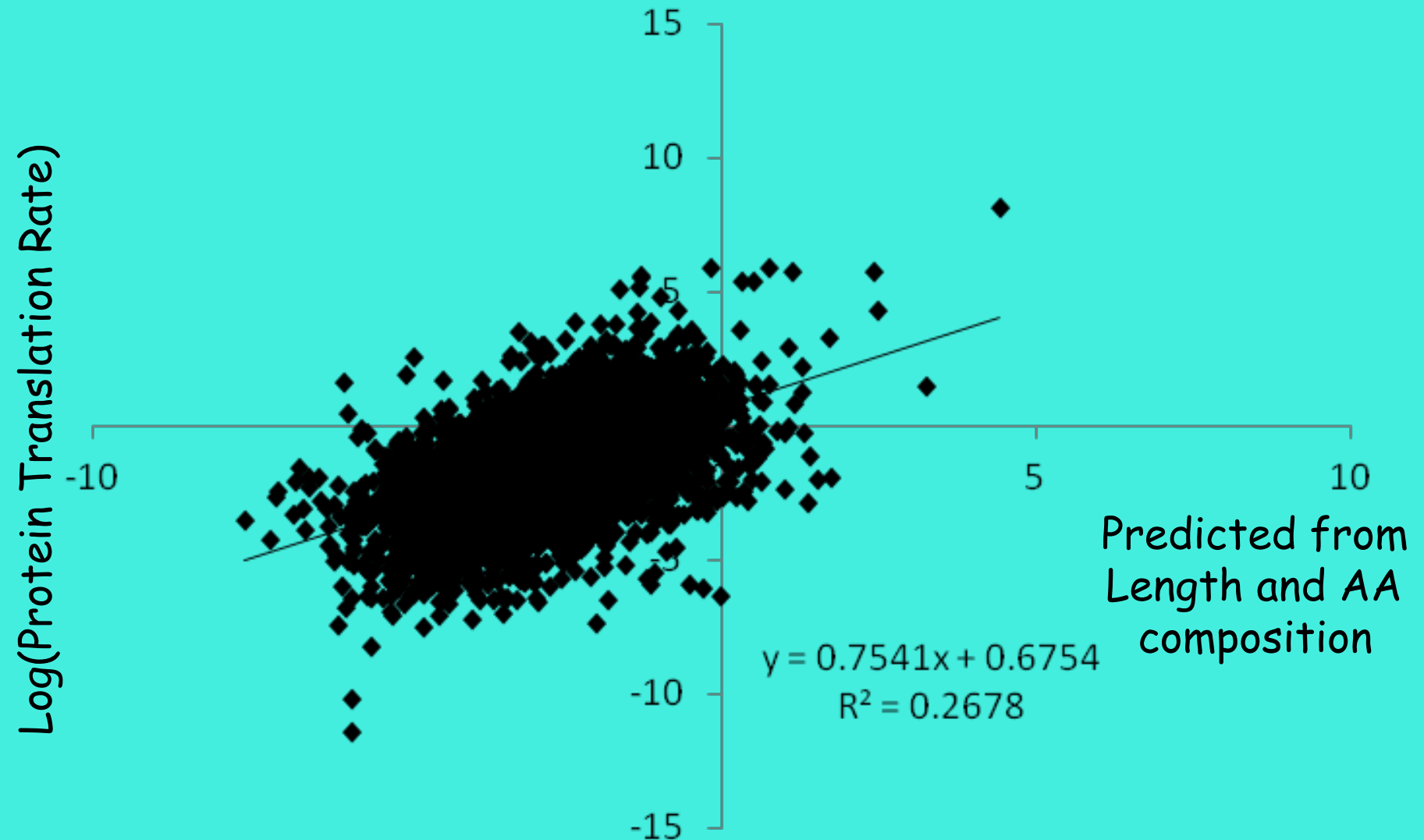


Differences between transcriptomics and proteomics

- The number of protein molecules per mRNA: 1:1 to 1000:1

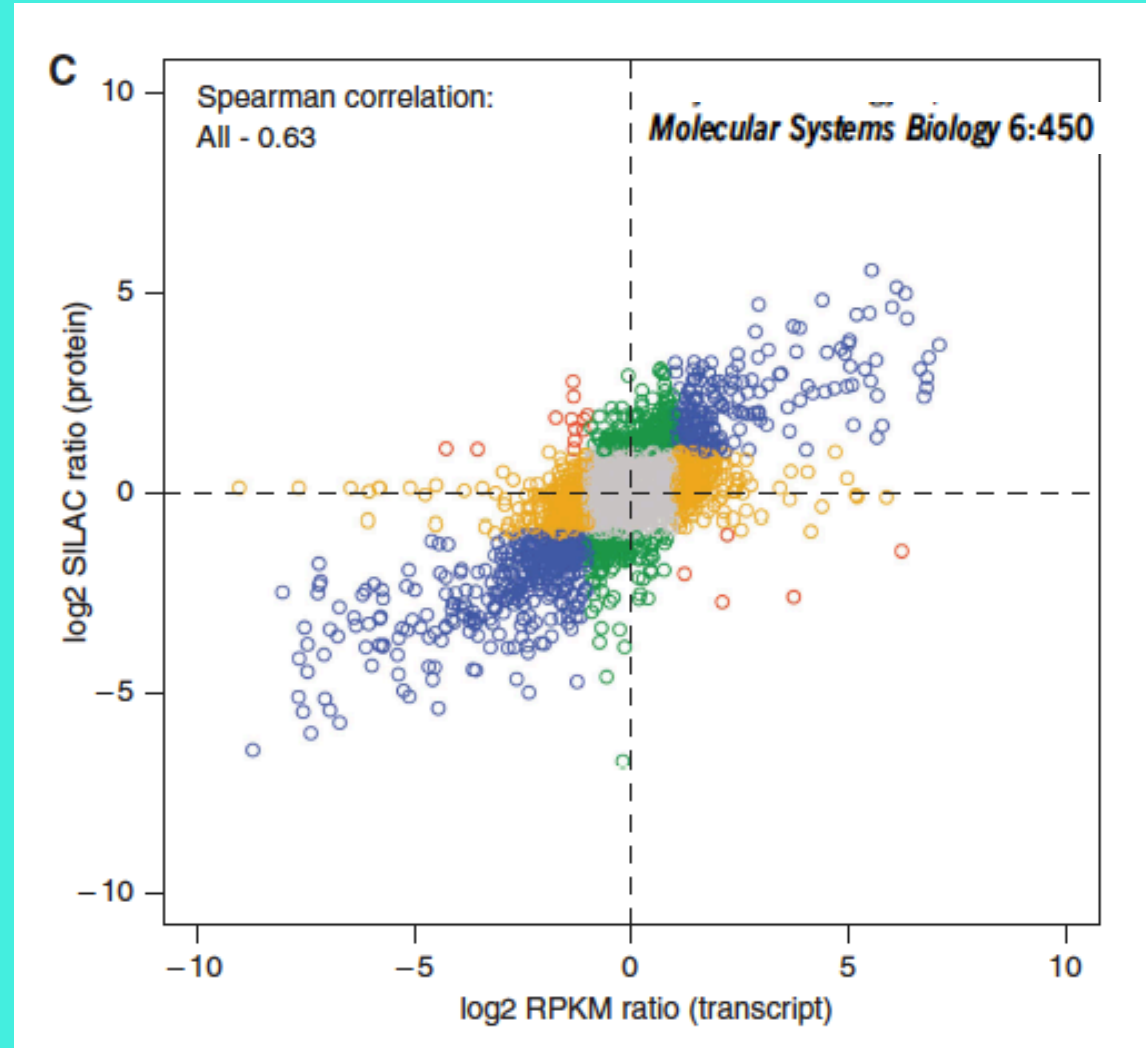


Combined Predictions - Length and AA Score



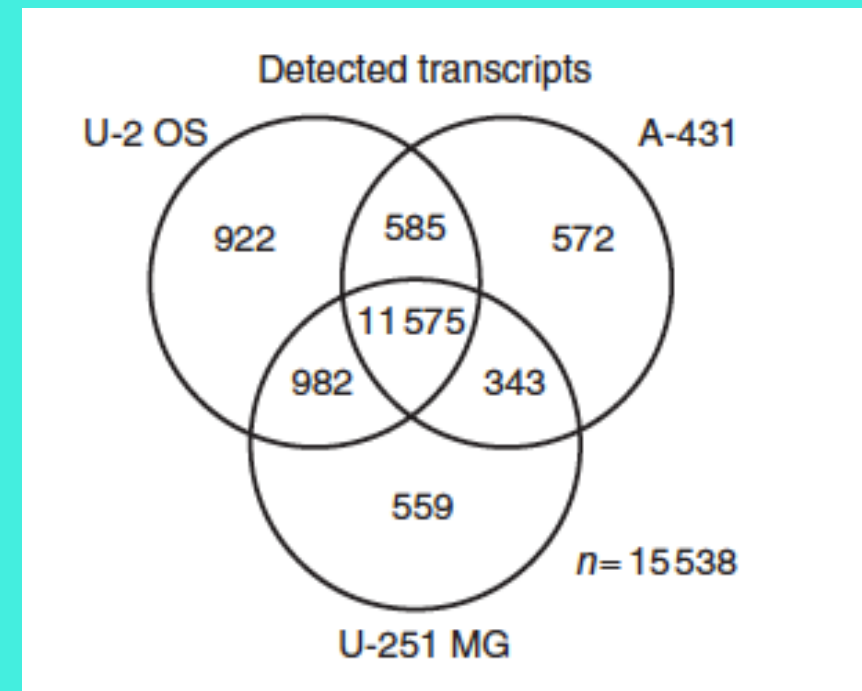
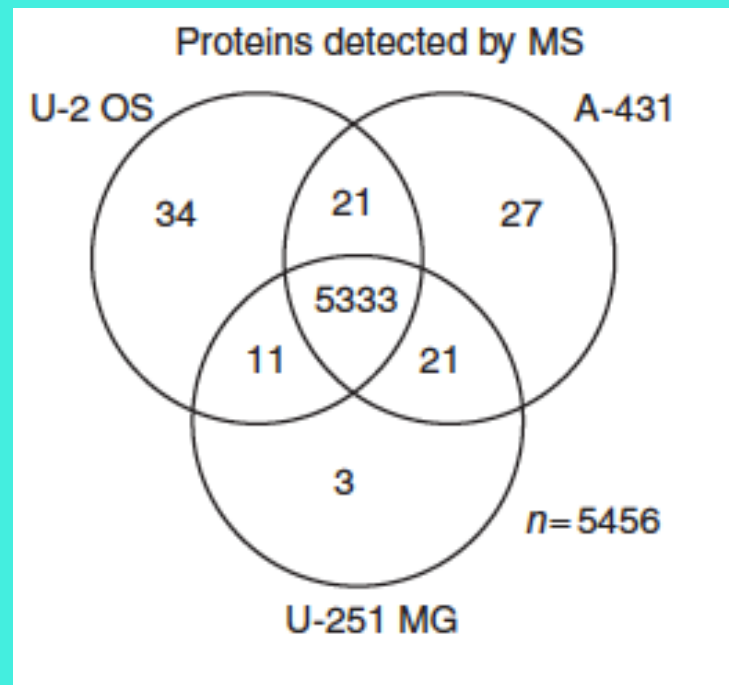
Other factors contribute to translation rate!

- mRNA abundances predict ca. 40% of the protein abundance, but $\log(\text{Ratio})$ for mRNA predict >60% of $\log(\text{Ratio})$ for proteins



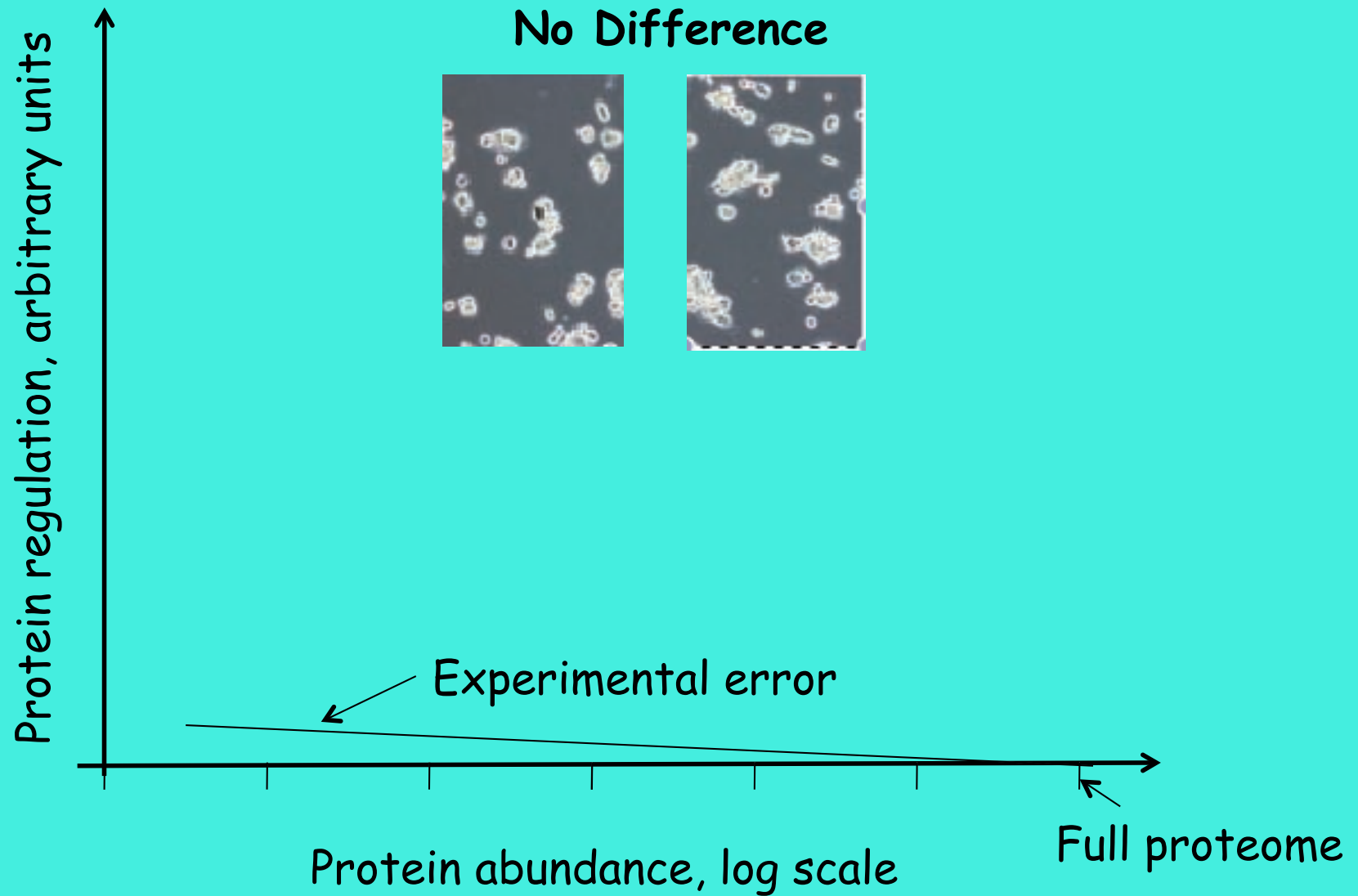
mRNA data need to be complemented by Proteomics data

In three different cell lines, practically all expressed genes (and proteins) are shared

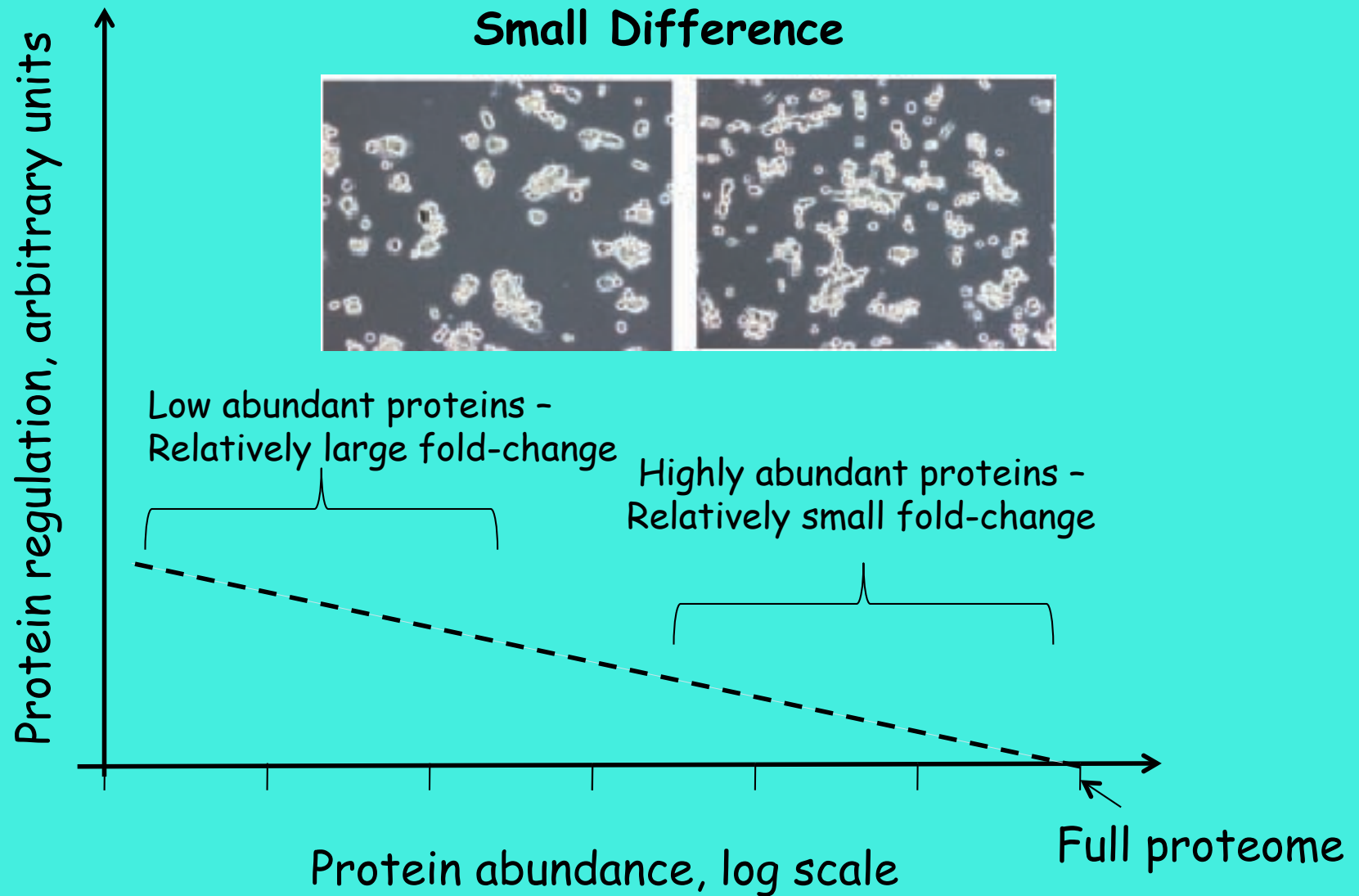


Same proteins are expressed in every cell type, but with different abundances

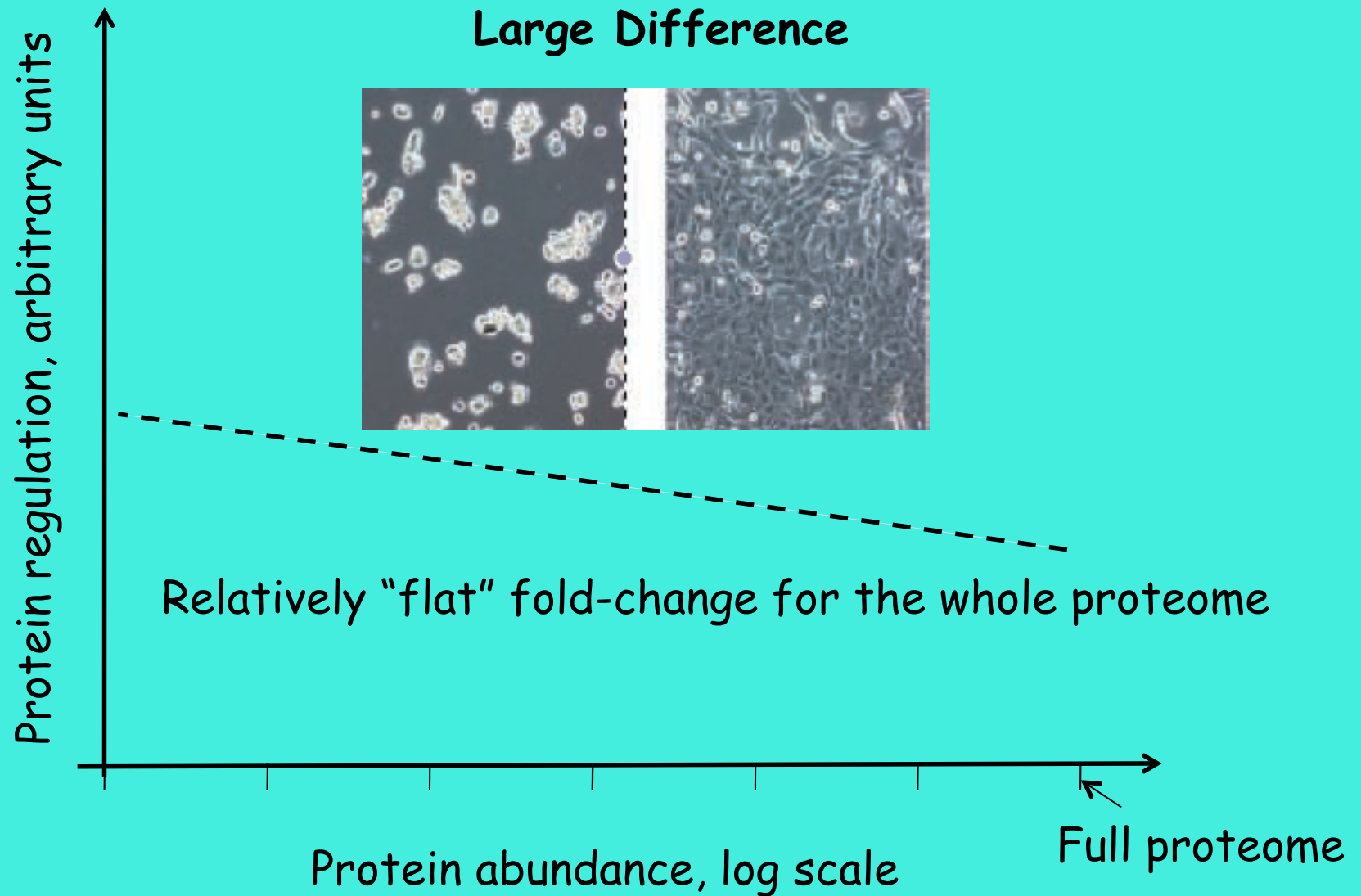
How does protein regulation depend upon protein abundance?



How does protein regulation depend upon protein abundance?



How does protein regulation depend upon protein abundance?

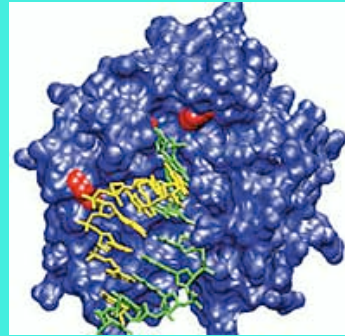


SUMMARY

- Transcriptomics provides large (95%) coverage of expressed genes, but it explains, at best, only 40% of the $\log(\text{Ratio})$ of protein abundances.
- Proteomics gives lower coverage (50% or less) by expressed proteins, but false discovery rate is only 1%
- For small changes in the proteome (e.g. early stages in time course), **deep** proteomics is advantageous, as proteins with significant fold-change are those of low-abundance
- For large changes in the proteome (e.g. cell type differentiation), even limited depth proteomics can provide specific fingerprint of cellular state, as protein regulation is largely independent upon abundance

Data Processing in Proteomics

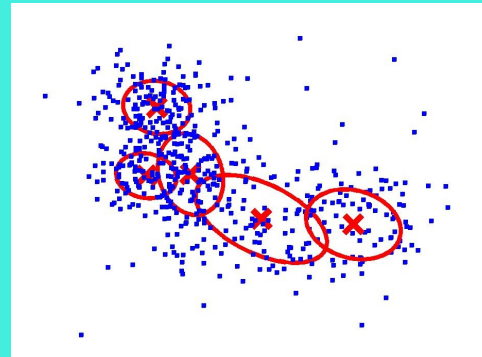
Reductionist Molecular Biology:



“golden bullet”

- detailed interactions, modifications, mechanisms
- lack of total picture

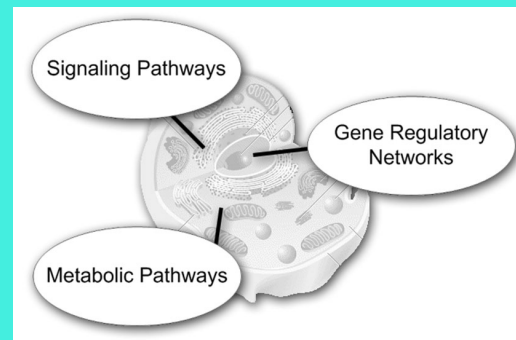
Statistical Approach:



Ad hoc, empirical model

- You get what you see
- Prediction, accuracy
- No explanation

Pathway Biology:



Global model

- prediction based on known pathways
- unknown accuracy
- do pathways exist?...

Protein Identification by Tandem Mass Spectrometry

Protein sequence

ILNKPEDETHLEAQPTDASAQFIRNLQISNE
DLSKEPSISREDLISKEQIVIRSSRQPQSQNPK
LPLSILKEKHLRNATLGSEETTEHTPSDASTT
EGKLMELGHKIMRNLENTVKETIKYLSLF
SHAFEVVK

Enzymatic
→
digest

Tryptic peptides

EDLISK
EQIVIR
LPLSILK
NLENTVK
LMELGHK
QPQSQNPK
NLQISNEDLSK
SLFSHAFAEVVK
NATLGSEETTEHTPSDASTTEGK
ILNKPEDETHLEAQPTDASAQFIR

Tandem Mass Spectrometry (MS/MS)

Tryptic peptide

NLENTVK

MS/MS

Fragmentation



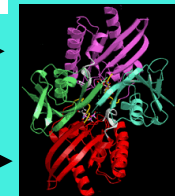
Molecular mass: 817.44

Fragment masses

232.17
346.22
388.20
444.28
484.33
511.37
555.40
623.45
666.44
712.52

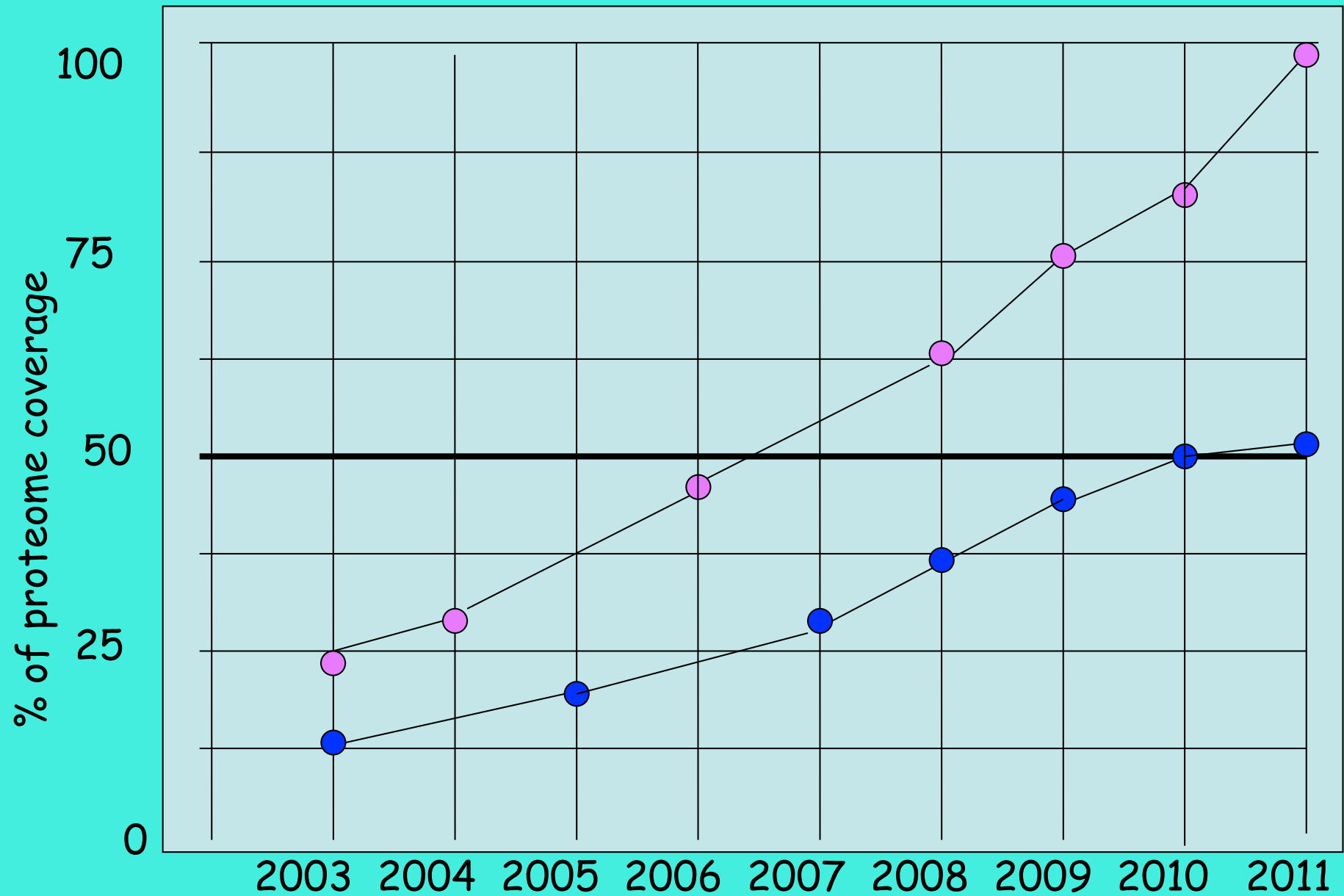


Your Peptide/
protein is this:



Score = 77

"Deep" vs "Top" Proteomics

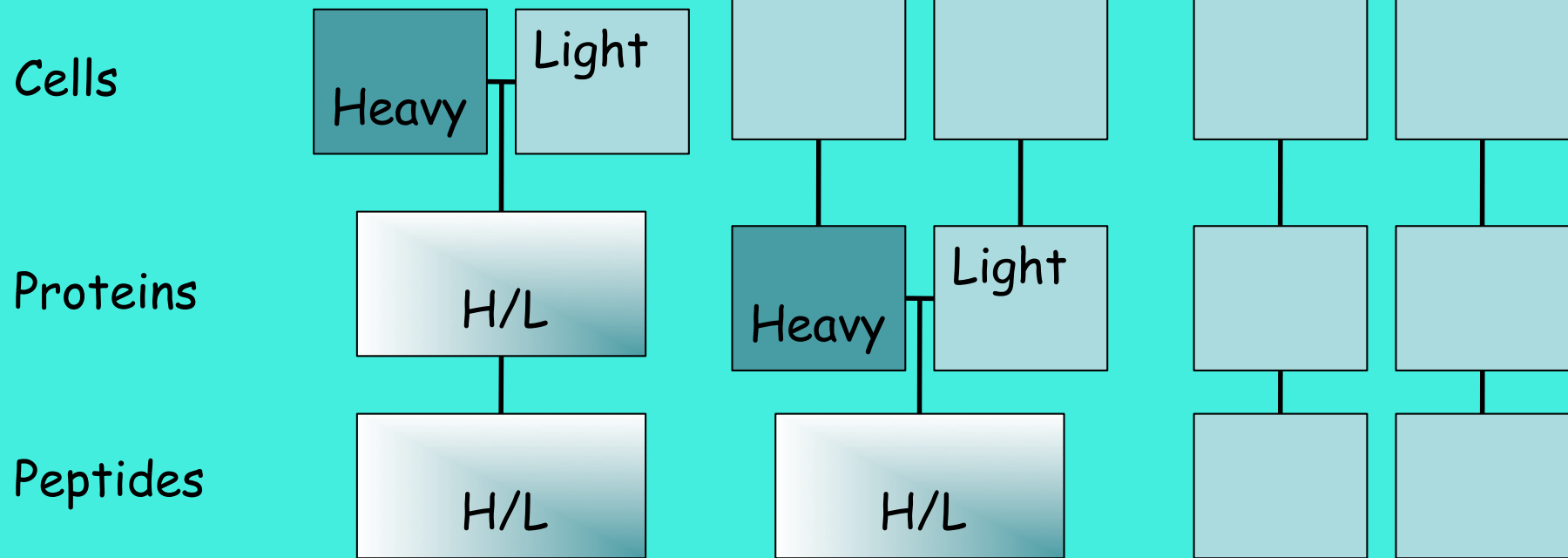


MS-based quantitative discovery approaches

SILAC

TMT, iTRAQ

Label-free



Biology

?

0

0

Extraction

0

x

x

Digestion

0

x

x

Tissues, bodily fluids

N/A

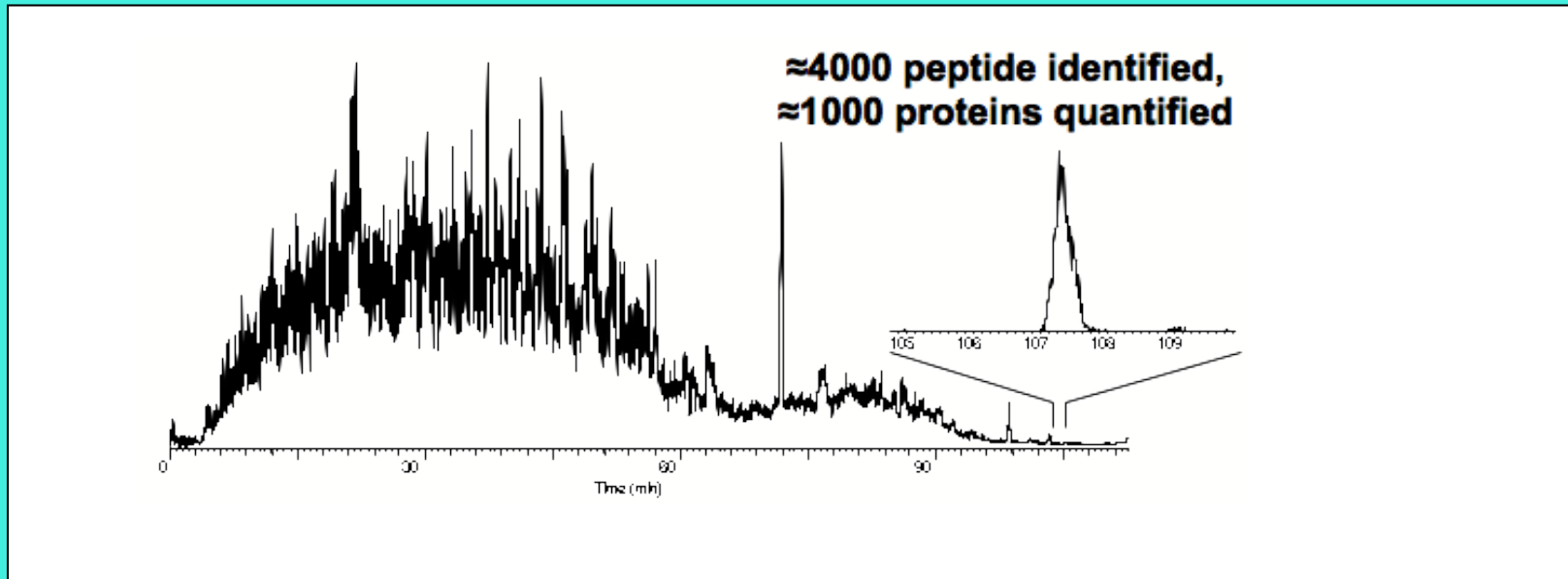
$N \leq 8$

N-any

Deep proteomics

Top proteomics

Top Proteomics



- 'Top proteome' : 1500-3000 proteins, 5000-9000 peptides
- No protein separation
- No peptide separation (on-line reverse-phase LC only)
- Single LC/MS experiment, 0.5-2.0 h long

Quanti 2.4 - February 2011 (2.5 - Feb 2012)

Quanti - ver. 2.4.0.0

DAT in: ...

db3: ...

Input Raw Files: Processing iLog: Reporting

+D:\Ft Data\Data Samples\5FU-9 dec\CMV_9dec010_NV_5fu_rko_B_0.raw
+D:\Ft Data\Data Samples\5FU-9 dec\CMV_9dec010_NV_5fu_rko_A_0.raw
+D:\Ft Data\Data Samples\5FU-9 dec\CMV_9dec010_NV_5fu_rko_A_30min.raw
+D:\Ft Data\Data Samples\5FU-9 dec\CMV_9dec010_NV_5fu_rko_C_30min.raw
+D:\Ft Data\Data Samples\5FU-9 dec\CMV_9dec010_NV_5fu_rko_B_30min.raw
+D:\Ft Data\Data Samples\5FU-9 dec\CMV_9dec010_NV_5fu_rko_C_0.raw
D:\Ft Data\Data Samples\5FU-9 dec\CMV_9dec010_NV_5fu_rko_A_1h.raw
D:\Ft Data\Data Samples\5FU-9 dec\CMV_9dec010_NV_5fu_rko_B_1h.raw
D:\Ft Data\Data Samples\5FU-9 dec\CMV_9dec010_NV_5fu_rko_C_1h.raw

Add... Delete

Output path: ...

Filter options

Mascot Score >= RT ± %:

Mass ± ppm: RT ± min:

Min peptides per protein:

Misc. Options

Charge Deconvolution
 Use best mascot peptide
 Quantify all mascot entries (.all file)

Reference file options

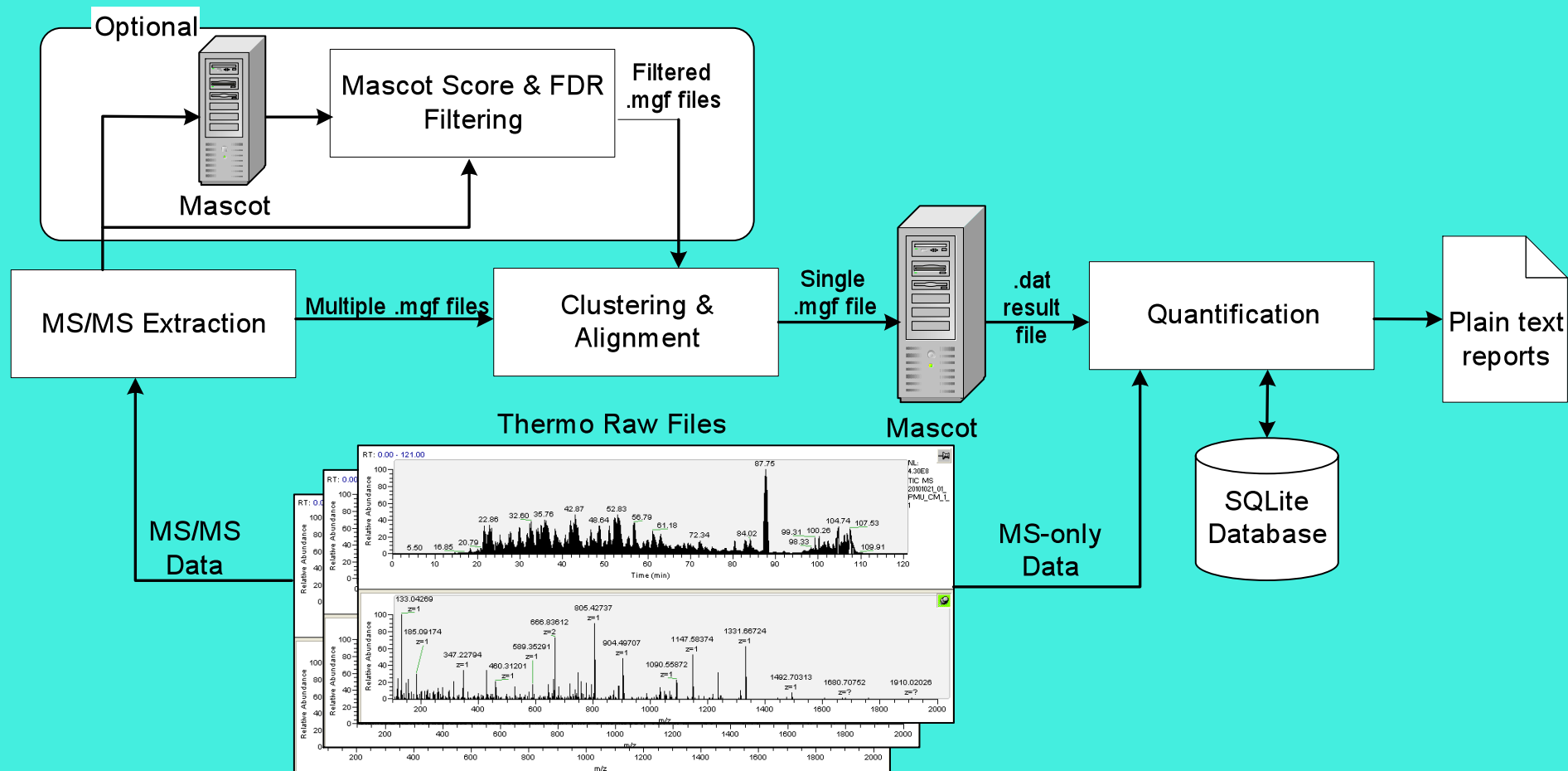
Reference file: ▾

ESI correction interval: Peak Shape Filtering

Normalization Cut tails

Loading completed

Quanti workflow



Pathway Analysis

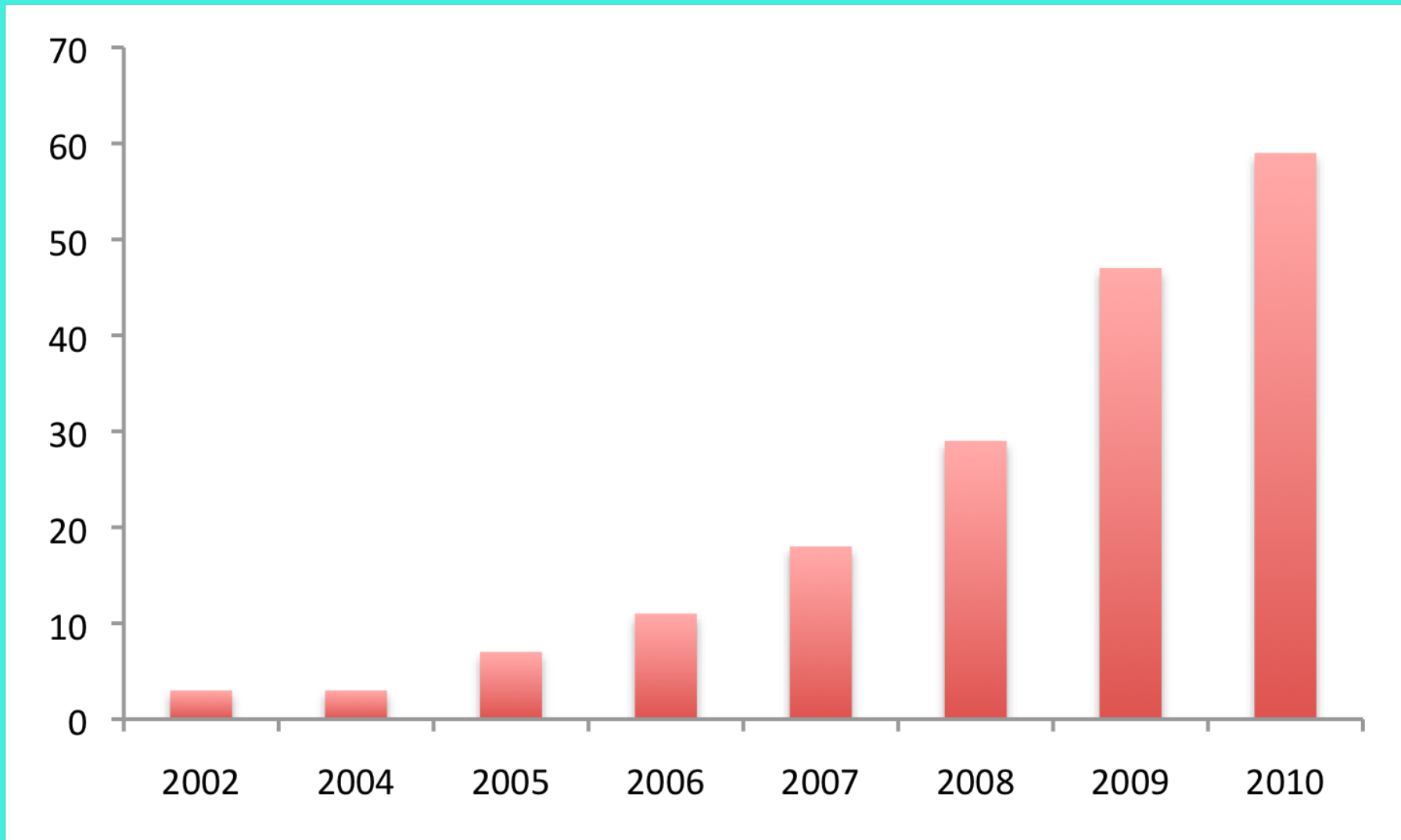
Disease Modeling

Drug Target Discovery

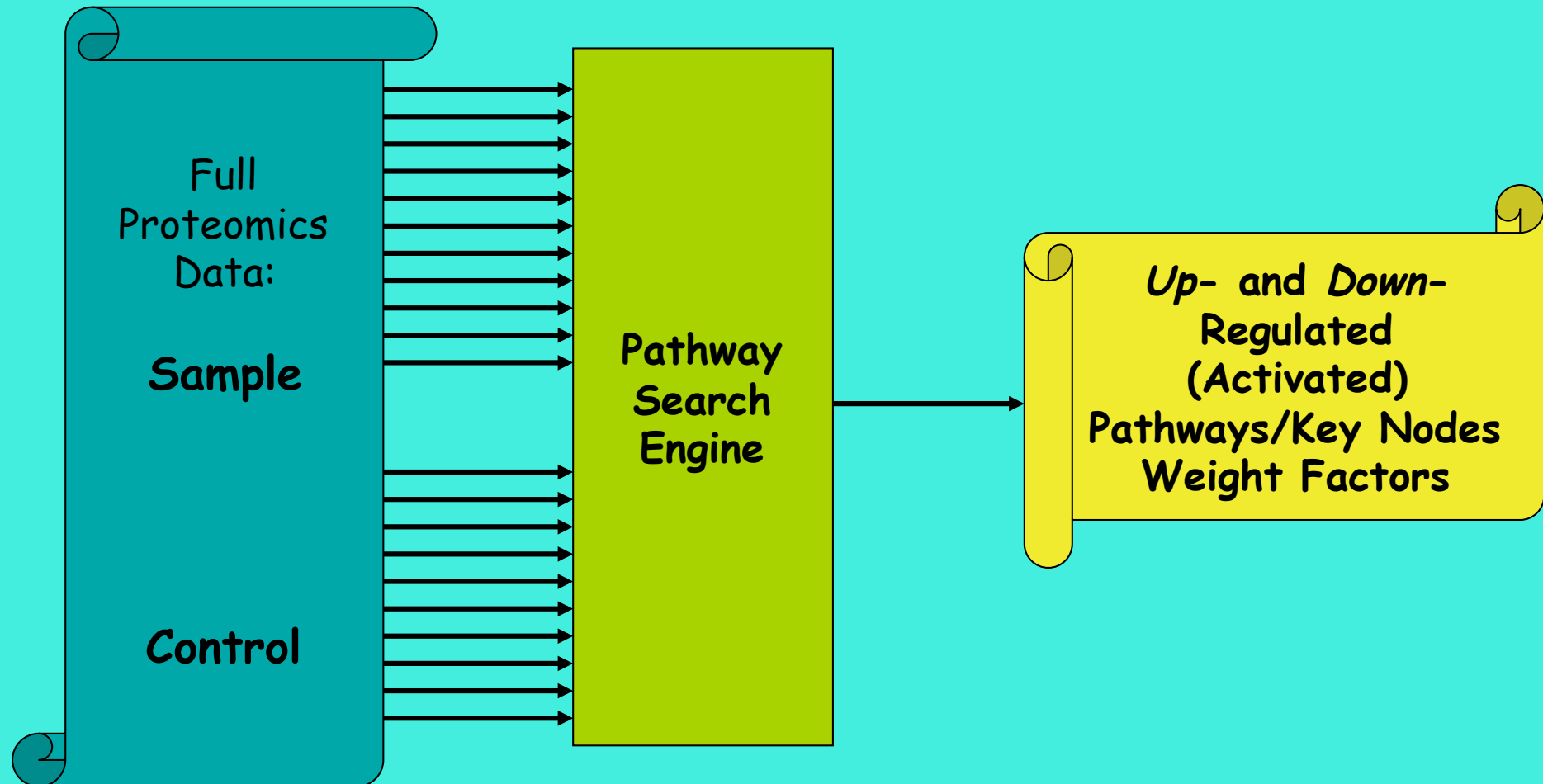
Patient Stratification

Establishing Drug Mechanism

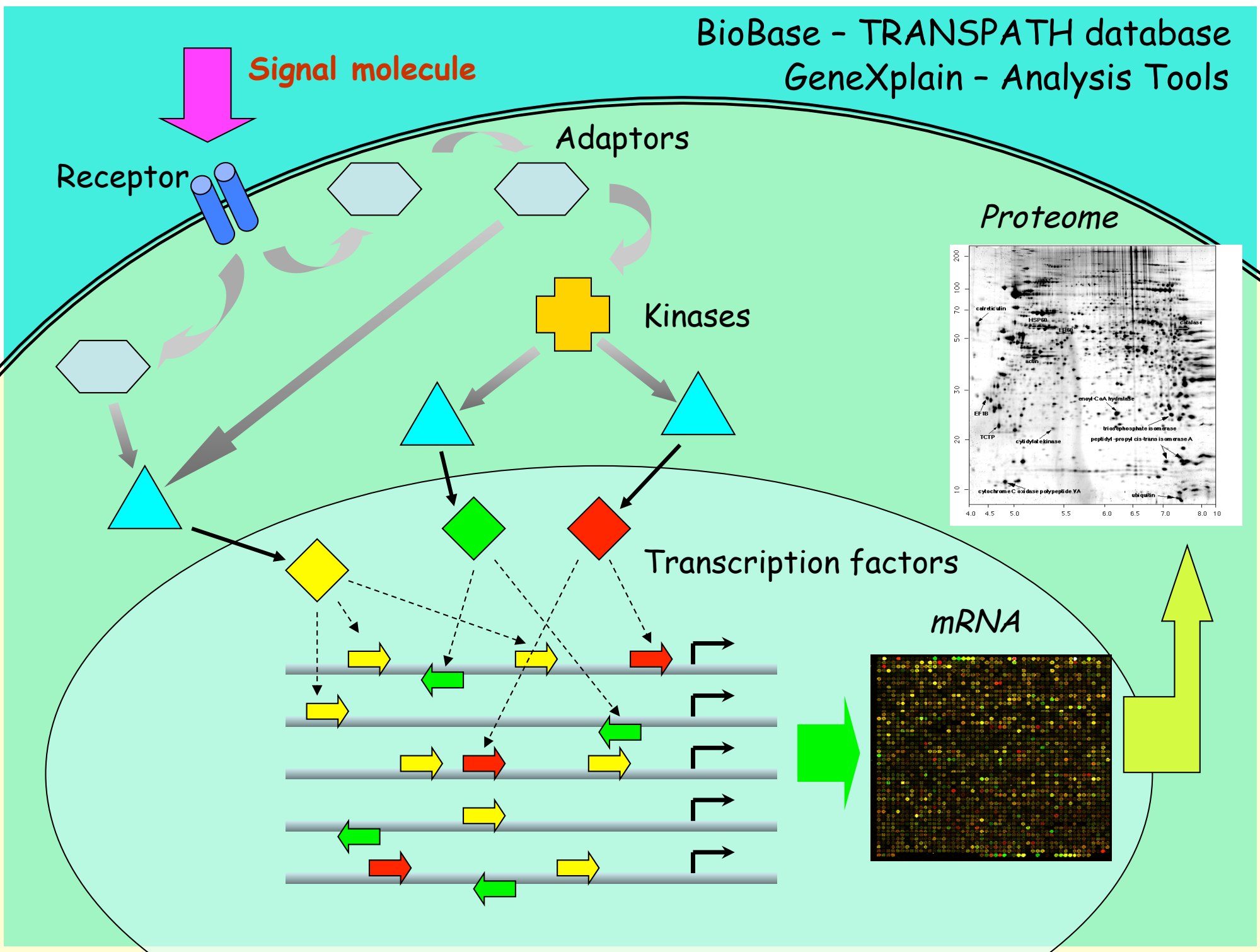
Pathway Analysis & Proteomics



Analytical Pathway Biology



Zubarev, R. A.; Nielsen, M. L.; Savitski, M. M.; Kel-Margoulis, O.; Wingender, E.; Kel, A.
Identification of dominant signaling pathways from proteomics expression data,
J. Proteomics, 2008, 1, 89-96.



Signal molecule

Receptor

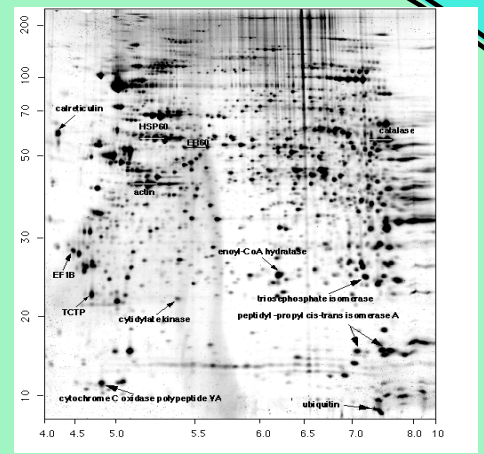
Adaptors

Kinases

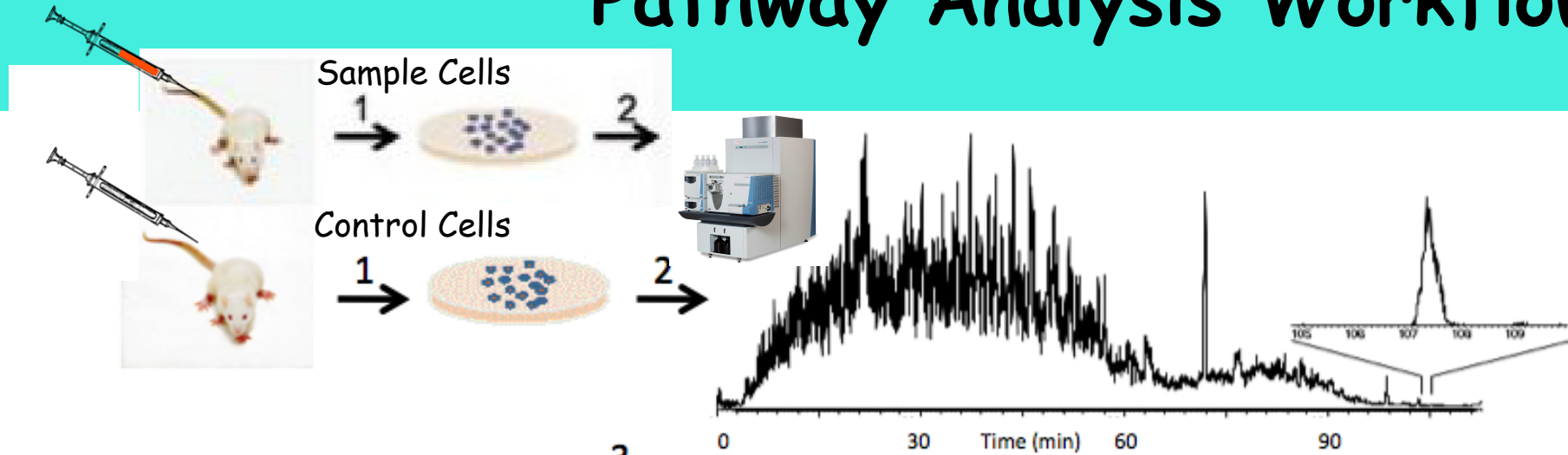
Transcription factors

mRNA

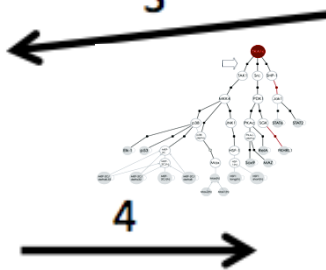
Proteome



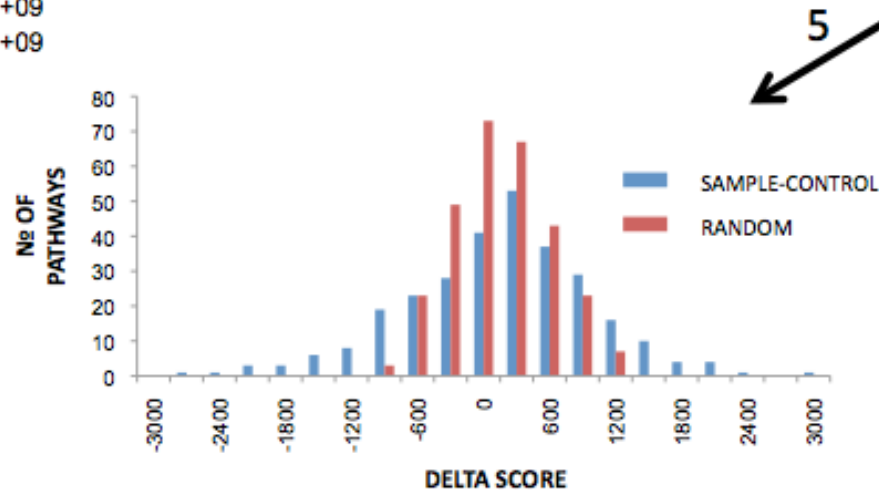
Pathway Analysis Workflow



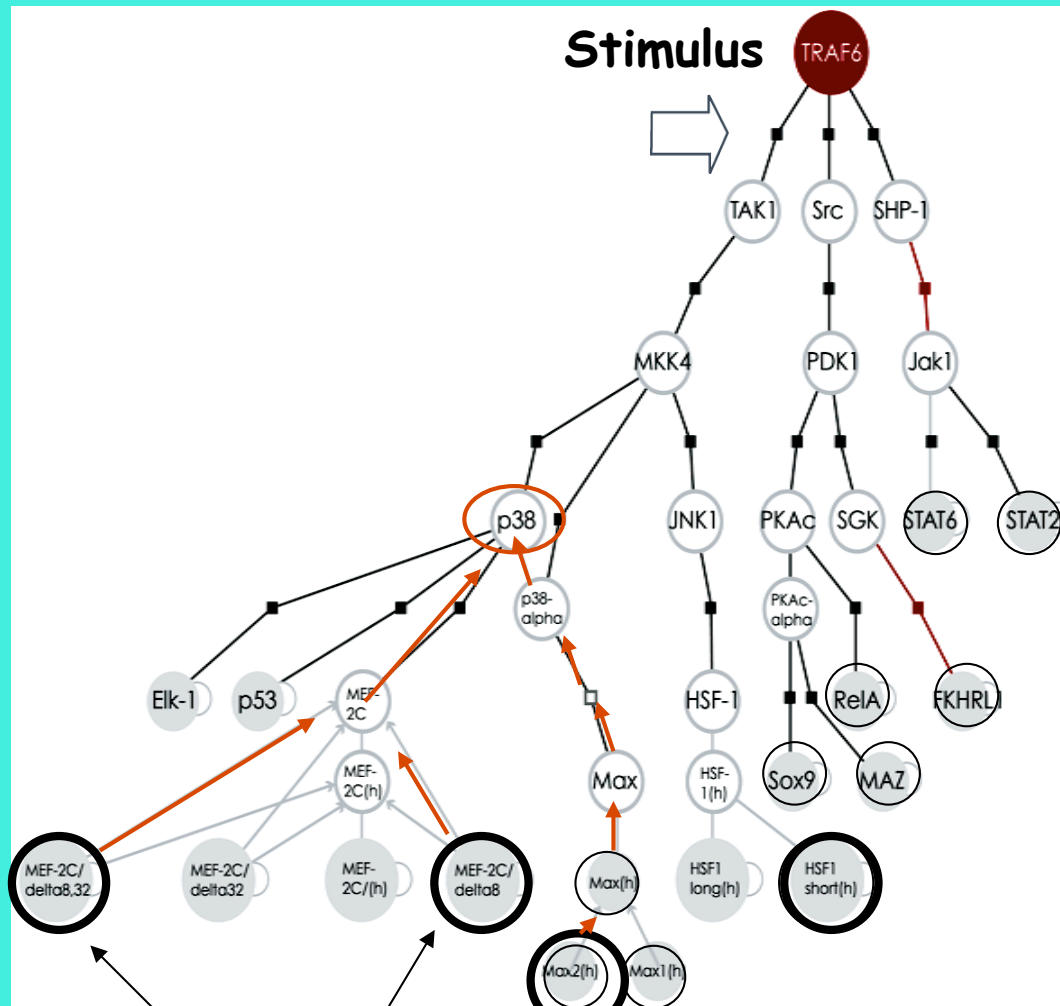
ACCESSION №	CONTROL	SAMPLE
IPI00131695	6.91E+08	3.71E+09
IPI00319992	4.06E+09	8.84E+08
IPI00653158	3.23E+08	2.84E+09
IPI00830313	3.86E+09	6.35E+08
IPI00468481	1.28E+09	2.21E+09
IPI00312058	4.21E+09	2.61E+09
IPI00116753	3.62E+09	3.87E+09
IPI00652371	8.01E+07	3.26E+09
IPI00880839	2.64E+09	2.96E+09
IPI00117914	3.66E+09	3.06E+09
IPI00122815	1.09E+09	1.54E+09



PATHWAY NAME	CONTROL	SAMPLE	DELTA
EGFpathway	5.97E+03	3.93E+03	-2.04E+03
JNKpathway	4.54E+03	2.76E+03	-1.78E+03
Faspathway	4.28E+03	2.03E+03	-2.25E+03
Caspasenetwork	3.97E+03	2.03E+03	-1.94E+03
E2Fnetwork	2.65E+03	2.02E+03	-6.26E+02
p53pathway	1.54E+03	2.02E+03	4.78E+02
stress-associatedpathways	1.53E+03	1.51E+03	-2.03E+01
insulinpathway	1.22E+03	1.33E+03	1.09E+02
T-cellantigenreceptorpathway	9.95E+01	1.15E+01	-8.80E+01



KeyNode-Mediated Analysis: Upstream

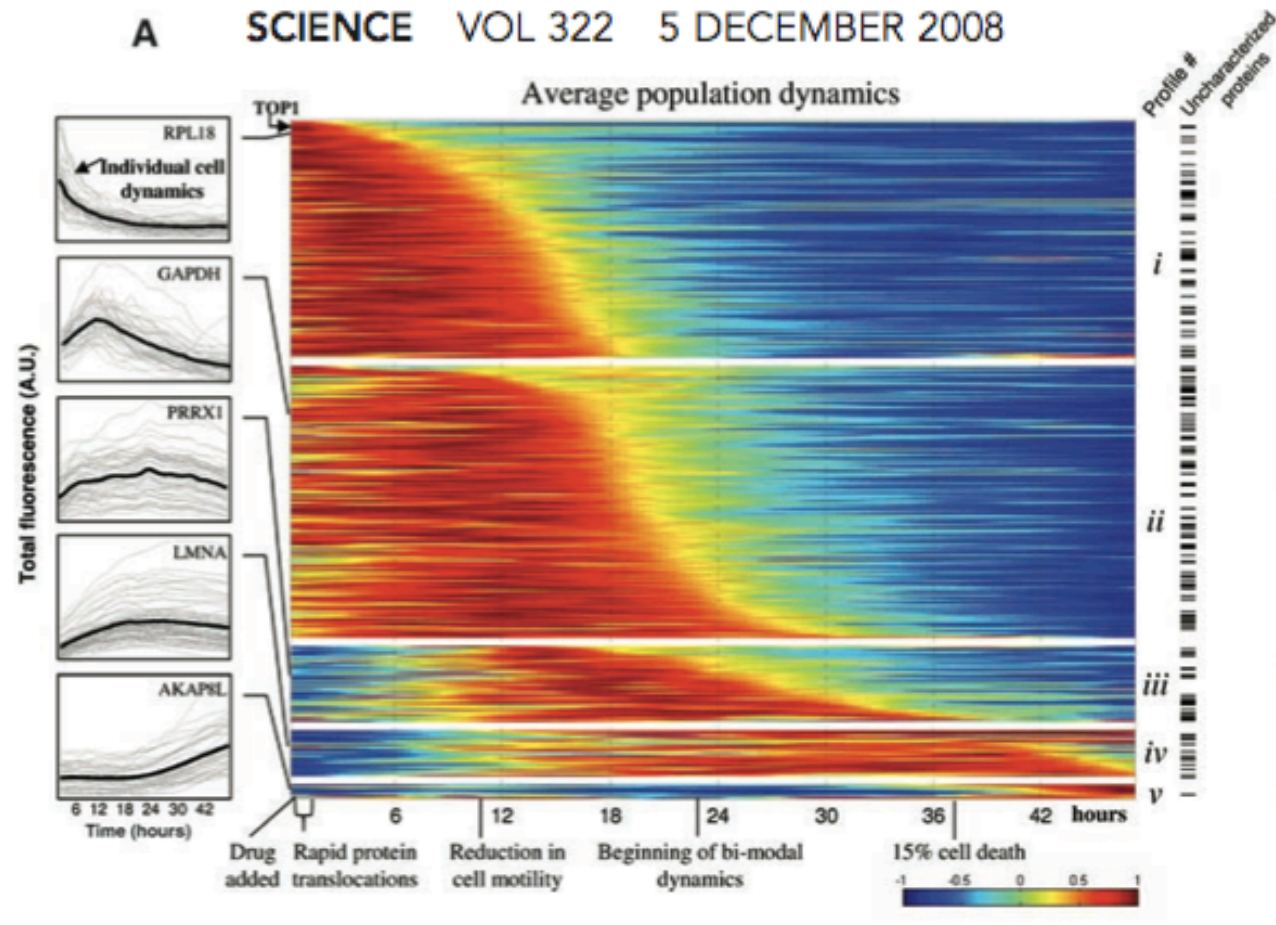


Proteins
Observed

Score

- KeyNode₁ 3050
- KeyNode₂ 2987
- KeyNode₃ 2073
- ...
- KeyNode_N 25

Pathway score:
 $\Sigma(\text{keynode score})$



DYNAMIC PROTEOMICS APPROACH

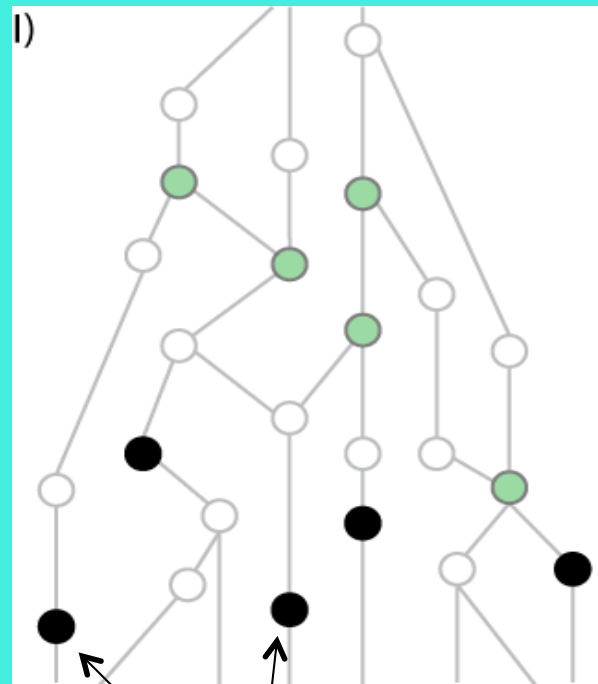
for drug target identification:

- by the **speed** of change (1 h), 10% selection
- by the total change in 48 h, 10% selection

Overall: top 3% (35 proteins)

Pathway Analysis of Dynamic Proteomics Data

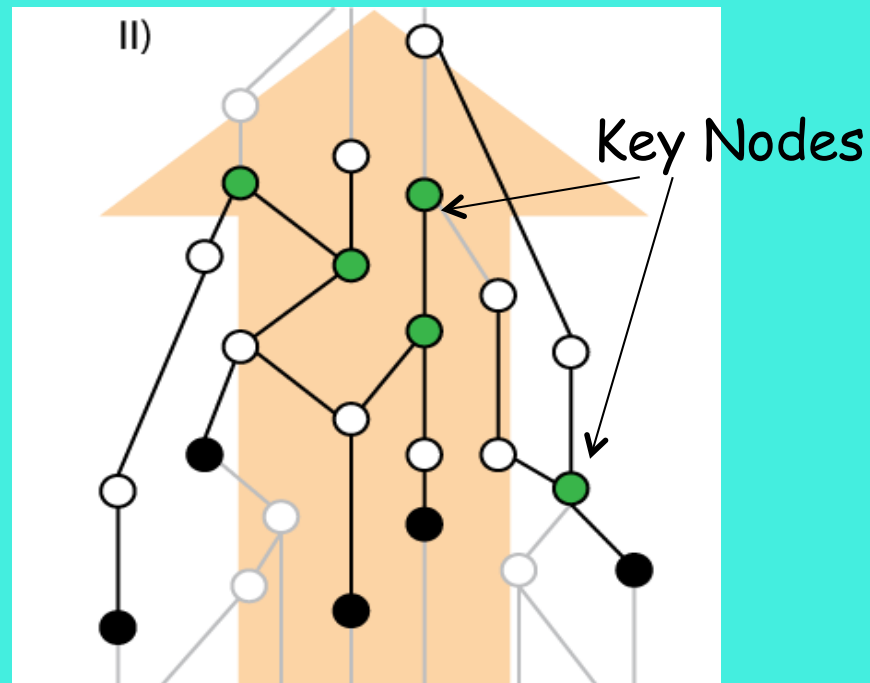
I) Protein mapping on Pathways



Proteins from
input list

Pathway Analysis of Dynamic Proteomics Data

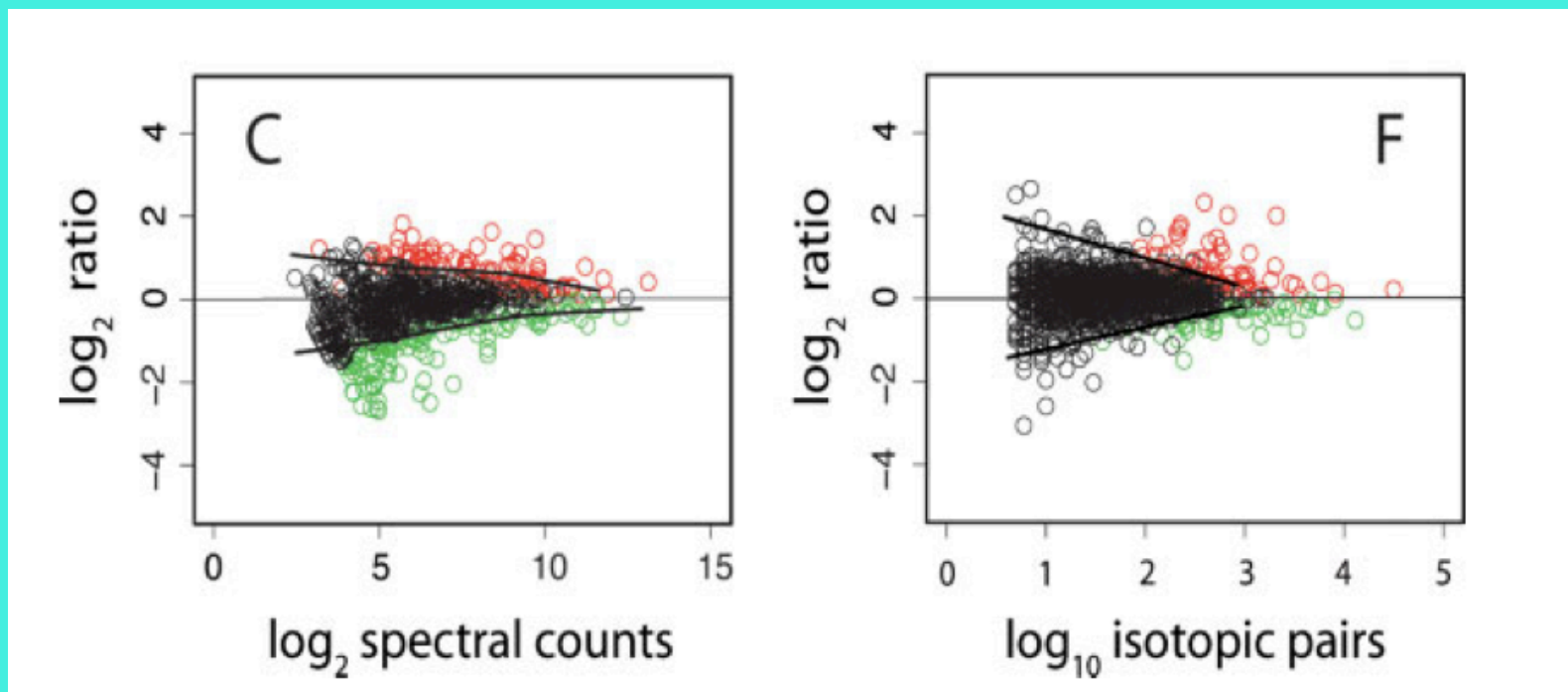
- Upstream Search:
- for Speed, 0-60 min
 - for Magnitude, 0-2800 min



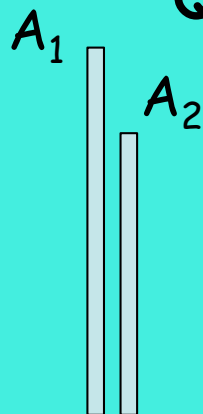
$$\text{KN Scoring: } \Delta S = (S_A - S_B) * \log_2(S_A/S_B)$$

Top KN is selected: one for Speed, one for Magnitude

The threshold problem in proteomics



Hacket M. *Science, Marketing and Wishful Thinking in Quantitative Proteomics*, *Proteomics*, 8 (2008).

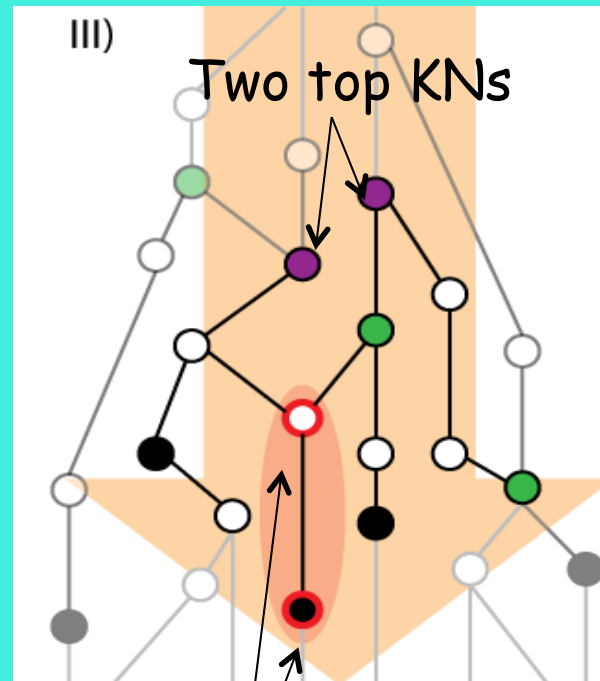


$$G = \text{Abs}(A_1 - A_2) \times \log_2(A_1 / A_2) \text{ [ppm]}$$

IF statistical fluctuations of protein abundances follow Poisson distribution, G -threshold is constant

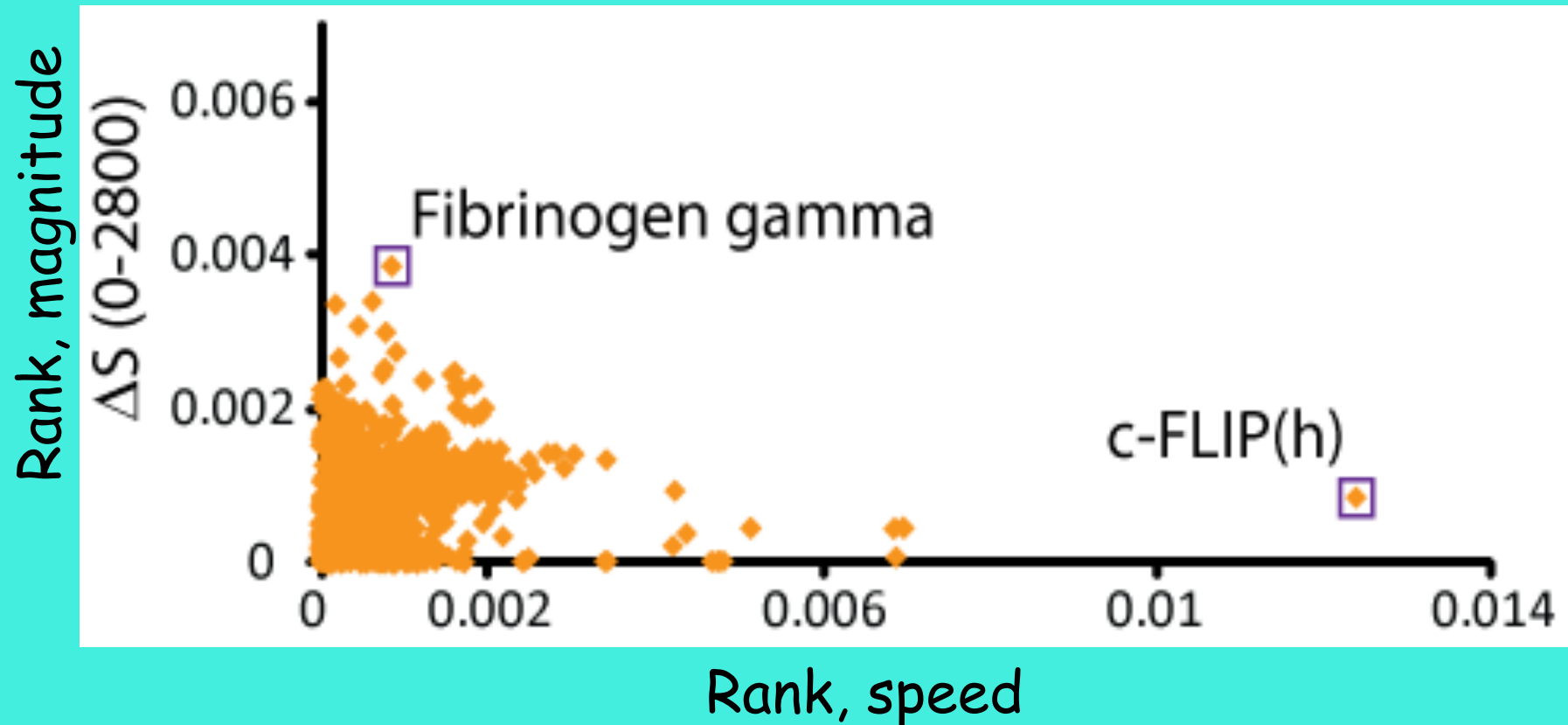
Pathway Analysis of Dynamic Proteomics Data

Downstream KN search



Overlapping
Molecules
= Drug Target Candidates

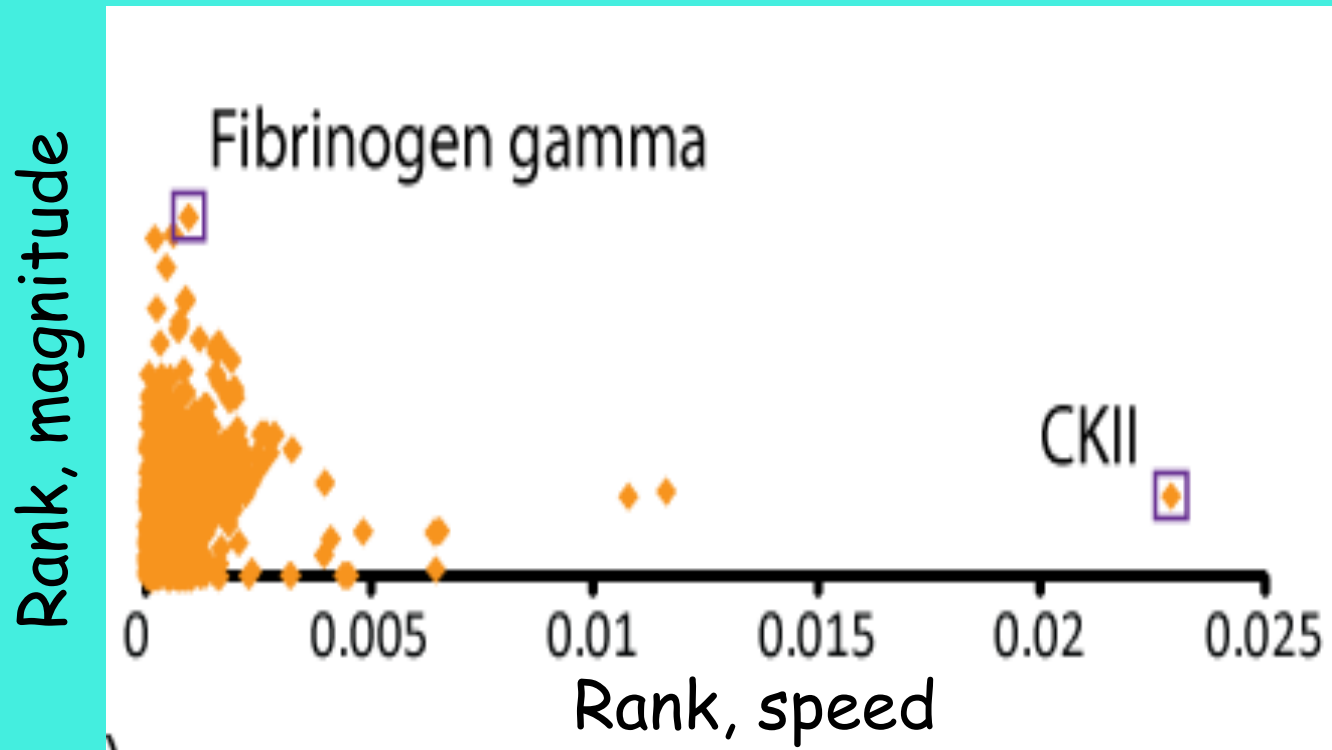
Identification of TOPI as *the* drug target from 812 proteins in the input list



Overlap of downstream lists from F_{gamma} , c-FLIP(h):
9 proteins, of which 2 from input list (known dynamics):

- TOPI, (speed + magnitude)-rank 228
- 26S proteasome, (speed+ magnitude)-rank 787

What if TOPI is *removed* from Input list?..



Overlap of downstream lists from F_{gamma} , c-FLIP(h):
4 proteins, none from the input list:

- TOPI
- CKII
- Two NR-related proteins

Take-home messages:

- Transcriptomics and proteomics overlap, but proteomics is "closer to action", and thus produces more relevant data
- Proteomics is currently limited in "depth" due to the large dynamic range of protein abundances, but technology moves forward fast, and the proteomics depth is increasing
- Correlation analysis provides first insight into the biological process, but pathway analysis is necessary to put the results in biological context
- Simple mapping of regulated proteins onto pathways ("direct mapping") often is insufficient;
- Upstream keynode analysis is superior over direct mapping
- Combining transcriptomics, proteomics and metabolomics data is the future goal of pathway analysis